

Intracellular delivery of CRISPR-Cas9 RNP for medical applications

211104

B4 Yuto Azumaya

① Introduction

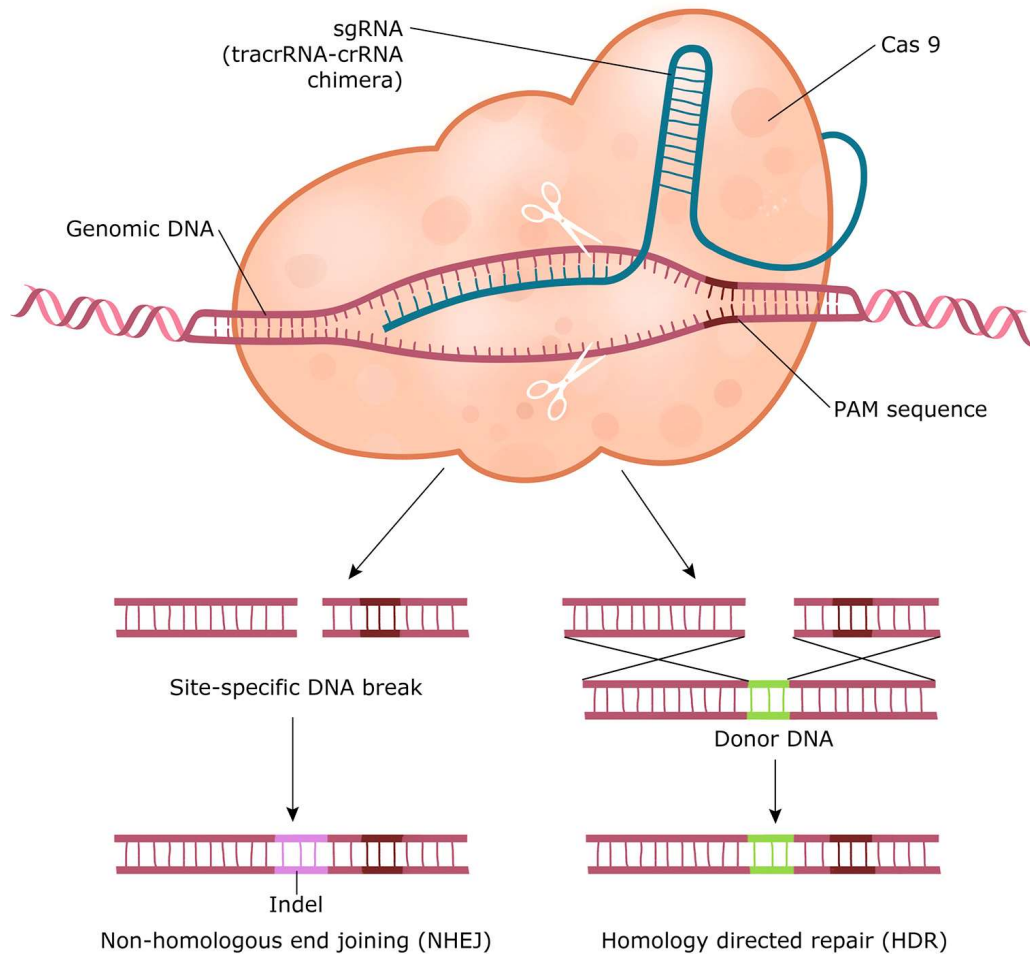
- 1) CRISPR/Cas9 system
- 2) Expectations for medical applications
- 3) other genome editing tools
- 4) Delivery form of CRISPR/Cas9 system
- 5) Investigational drugs using CRISPR-Cas9

② Methods of delivering CRISPR/Cas9 RNP

.

③ Delivery of systems that cause HDR with high yields

④ Summary



- A tool that can perform DNA editing
- Consists of Cas9 protein and gRNA
- The gRNA recognizes and cleaves the target sequence.
- Cleaved sequences are repaired mainly by end joining (NHEJ) or homologous recombination (HDR)
- NHEJ is prone to errors such as deletions and mutations → suitable for gene deletions
- HDR can introduce the desired sequence depending on the design of the donor DNA.

(Nataša Savić, Gerald Schwank, Translational Research, Volume 168, 2016, 15-21.)

Kim S, Broströmer E, Xing D, et al. *Science*. 2013;339(6121):816-819.
Ramirez-Phillips, A.C., Liu, D. *AAPS J* **23**, 80, 2021.

Diseases for which CRISPR-Cas9 system application is expected

- Diseases caused by genetic abnormalities: Editing pathogenic genes for radical treatment

Ex)

- Huntington's disease: nucleotide elongation disorder, CAG repeat expansion of the huntingtin gene
- Hemophilia: loss of clotting factors due to mutation
- Duchenne muscular dystrophy: generation of loss-of-function proteins by frameshifting
- sickle cell disease :deformation caused by a single Glu6Val mutation in the *HBB* gene etc.

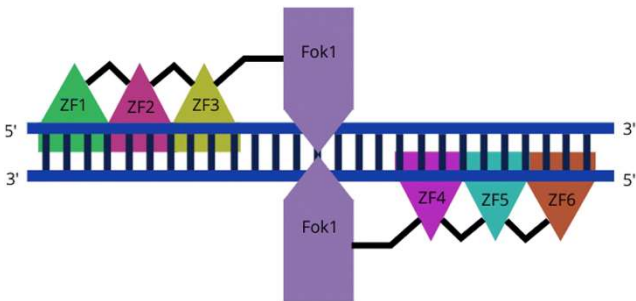
- Viral diseases: Disrupting viral genes to inhibit their function

Ex)

- HSV
- HIV
- HPV
- HBV

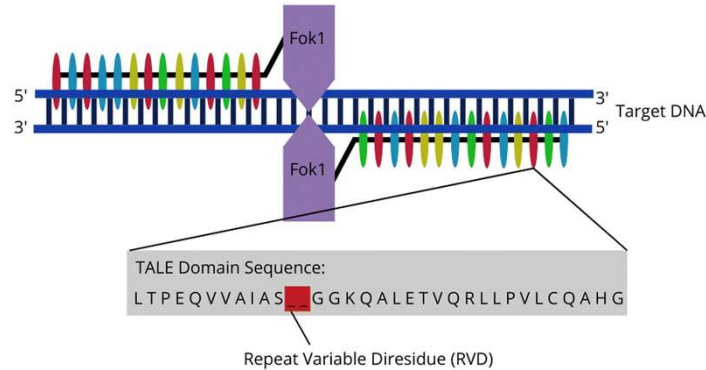
etc.

- Cancer: Treat cancer by deleting oncogenes or introducing tumor suppressor genes



ZFN

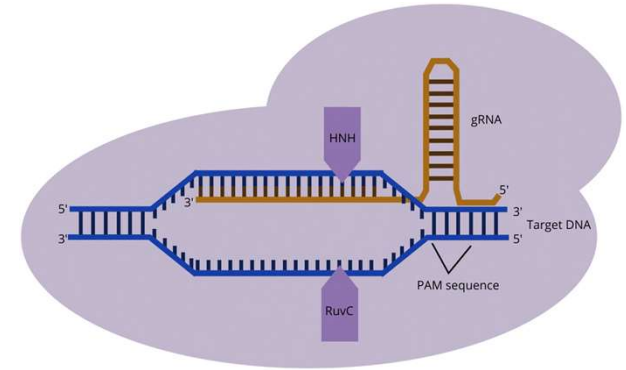
Fok-1 nuclease
+ Zinc finger protein
(recognizes the target sequence)



TALEN

Fok-1 nuclease
+TALE domain (Base is recognized by amino acid sequence of RVD variable region)

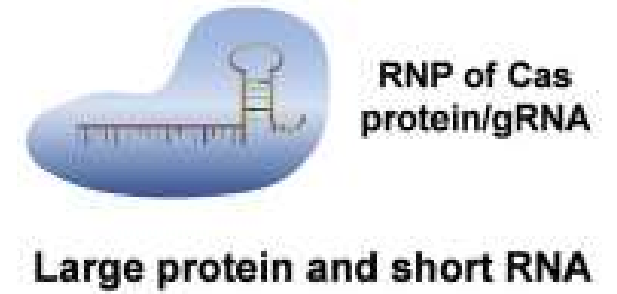
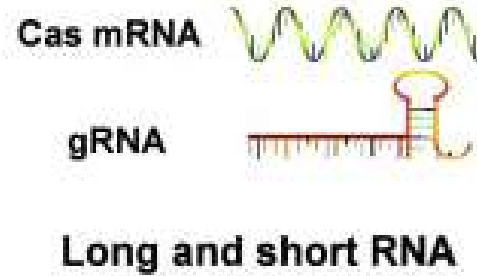
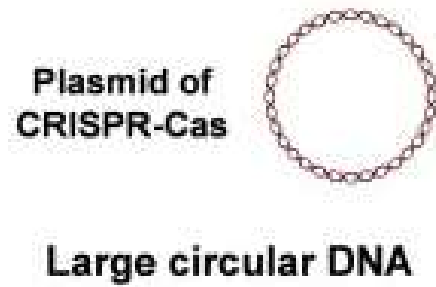
sequence-specific DNA-binding protein
→difficult and expensive to engineer.



CRISPR/Cas9

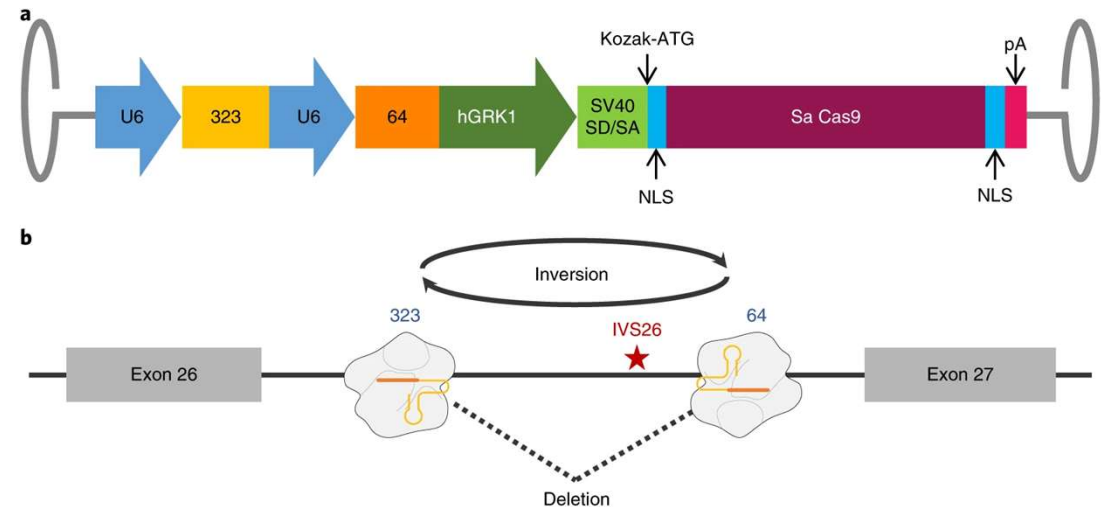
Cas9 nuclease
+ sgRNA(targeting genome)

- RNA : easily synthesizable & applicable to other genes by simply changing the RNA sequence.



EDIT-101

- Target disease: LCA10 (leber congenital amaurosis type 10)
- The IVS26 mutation in the CEP290 gene causes abnormal splicing, resulting in the addition of extra sequences to the mRNA → loss of function.
- Deletion or reversion of the mutation by cutting across the mutation site cures the splicing abnormality and allows normal CEP290 to be produced.
- Introduced by AAV in the form of pDNA
- Vitro: average productive editing rate (reverse + deletion) $16.6 \pm 6.5\%$
- Vivo: up to 28% in subretinal injection in cynomolgus monkeys (10% deletion thought to lead to visual recovery)
- Clinical trial (ongoing Phase 1, initial results): confirmed efficacy signal in 2 out of 3 patients in medium volume cohort

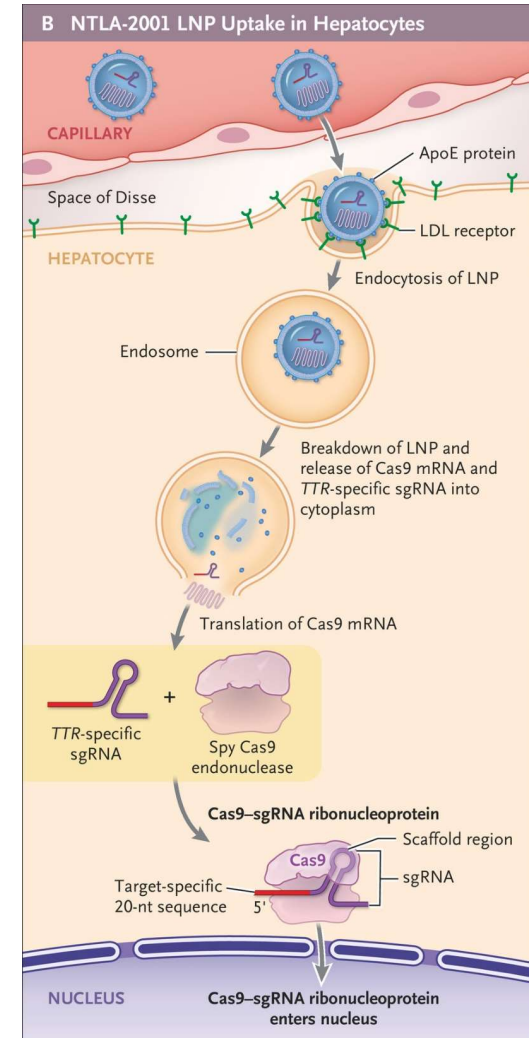


Maeder, M.L., et al. *Nat Med* 25, 229–233 (2019).

Investigational drugs using CRISPR-Cas9

- NTLA-2001
- Target disease: ATTR amyloidosis
- accumulation of misfolded transthyretin (TTR) protein → nerve damage or heart muscle disease
- TTR KO → decrease in the production of TTR

- lipid nanoparticle encapsulating Cas9 mRNA + sgRNA
- Introduced by intravenous infusion
- Vitro (primary human hepatocytes): Editing efficiency of more than 93.7%, decrease in the production of TTR of more than 95%, no off-target editing confirmed
- Vivo (cynomolgus monkeys): up to 73% editing efficiency, >94% serum TTR reduction, no off-target editing
- Clinical trial (on-going Phase 1, interim data): Sustained reduction in serum TTR protein concentration (up to 87%) was observed.





Large circular DNA

- stable
- simple to produce
- easy to deliver (ex vivo with physical methods)
(in vivo with viral vectors)
- Long-term Cas9 expression by transcription and translation
→ off-target DNA cleavage / host immune responses
- Possibility of insertional mutation



Long and short RNA

- limitation of the duration of Cas9 activity
→ Reduction of off-targets
- Does not cause insertional mutation
- Low stability of mRNA
- Requiring Translation

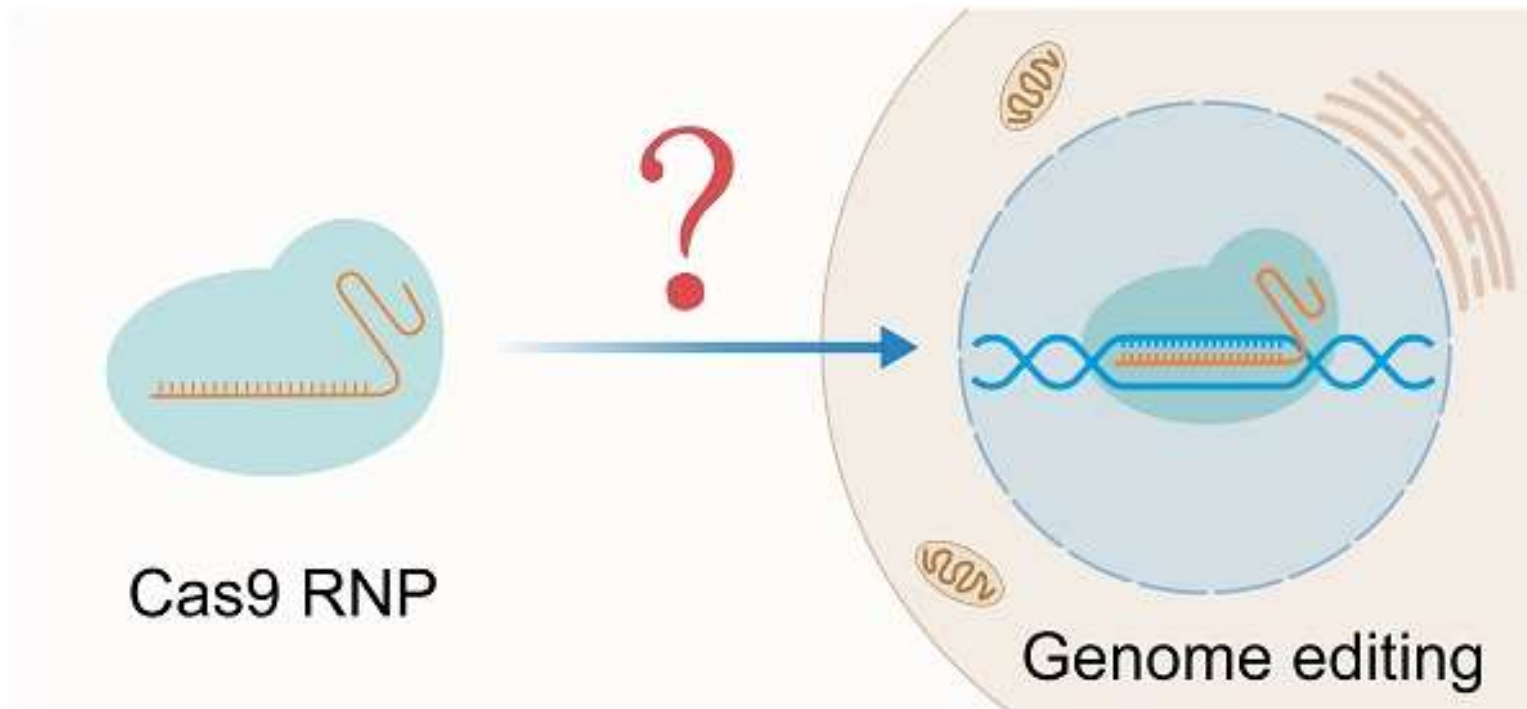


Large protein and short RNA

- No transcription and translation
→ Editing is possible even for cells with low transcriptional and translational activity.
- Easy to control the amount of transfection
- Quick editing
- High activity can be expected. (binding to sgRNA is not inhibited)
- Lowest possibility of off-target cleavage
- Reduction of immune response
- No insertional mutation

How to deliver?

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① Introduction

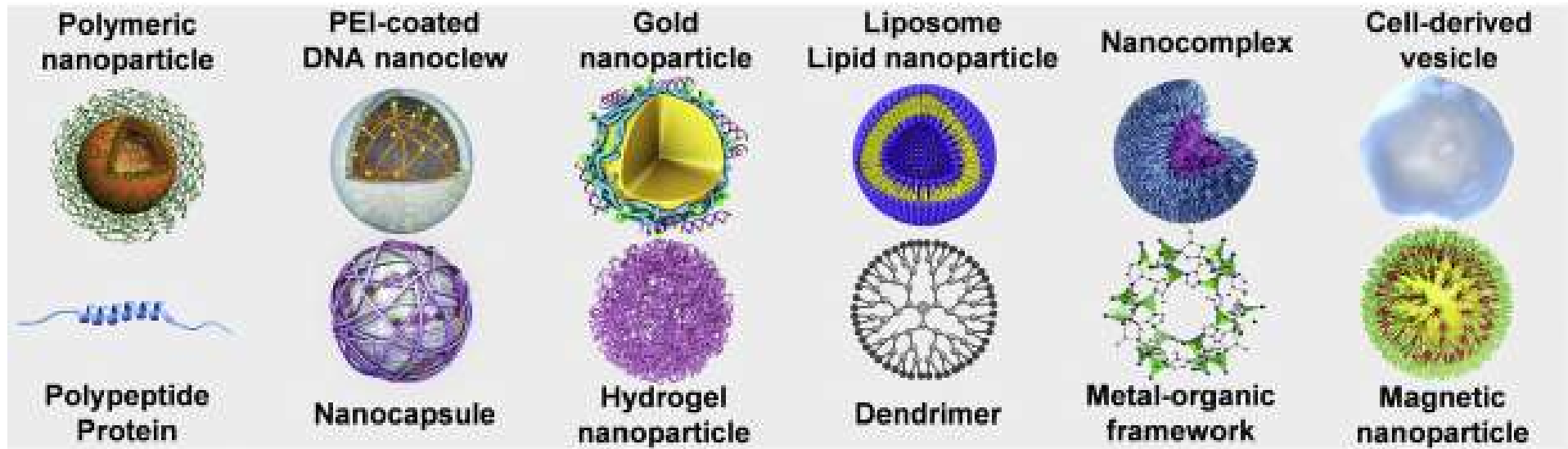
② Methods of delivering CRISPR/Cas9 RNP

- 1) General
- 2) Cationic substances commonly used for membrane permeation
- 3) Synthetic lipid nanoparticles
- 4) polymers
- 5) Inorganic nanoparticles
- 6) CPP conjugation
- 7) Biological Production

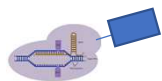
③ Delivery of systems that cause HDR with high yields

④ Summary

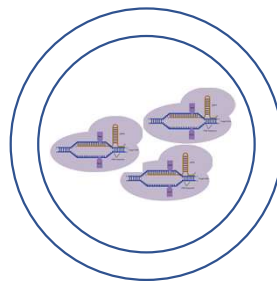
Various methods of delivering CRISPR/Cas9 RNP



Basic design concepts



or

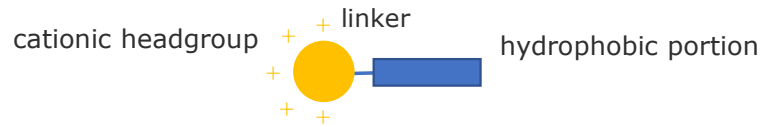


- ↓ Form nanoparticles
- ↓ Coating or attaching with membrane permeable molecules

- Membrane permeability: cationic lipids, PEI, CPP, etc.
- Nanoparticle formation: polymer, metal, DNA, etc.

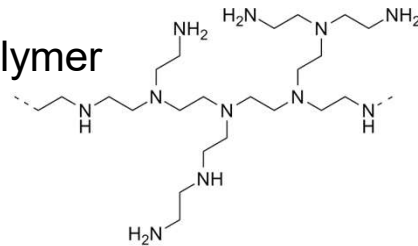
Cationic substances commonly used for membrane permeation

- Cationic lipids



- PEI

Polyethyleneimine, a polymer



- CPP

Peptides with cell membrane permeability
Many of them contain a large number of cationic residues.

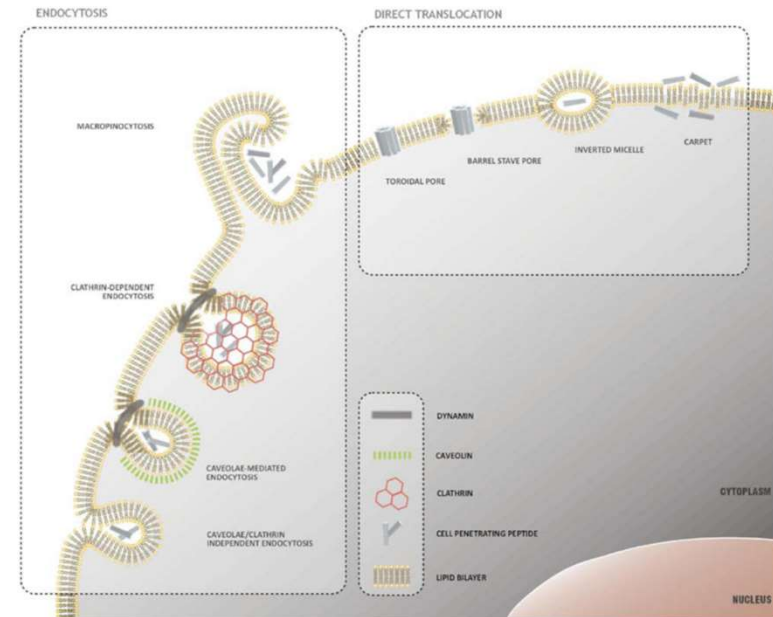
Table 1: Overview of some of the most commonly used CPPs describing their sequence, class and charge

Name	Amino acid sequence	CPP class	Charge
TAT	YGRKKRRQRRR	cationic	8
penetratin	RQIKWFGNRMK WKK	cationic	7
R9	RRRRRRRRR	cationic	9
MPG	GALFLGWLGAAGSTMGAPKKKKRV	amphipathic	24
Pep-1	KETWWTWTEWSQPKKRV	amphipathic	2
transportan-10	AGYLLGKINLKALAALAKKIL-amide	amphipathic	4
PepFect6	stearyl-AGYLLGK(c-TMQ)INLKALAALAKKIL	amphipathic	10
Bac7	RRIRPRPRLPRPRPRLPFPRPG	proline-rich	9

In both cases, uptake occurs by the following mechanisms

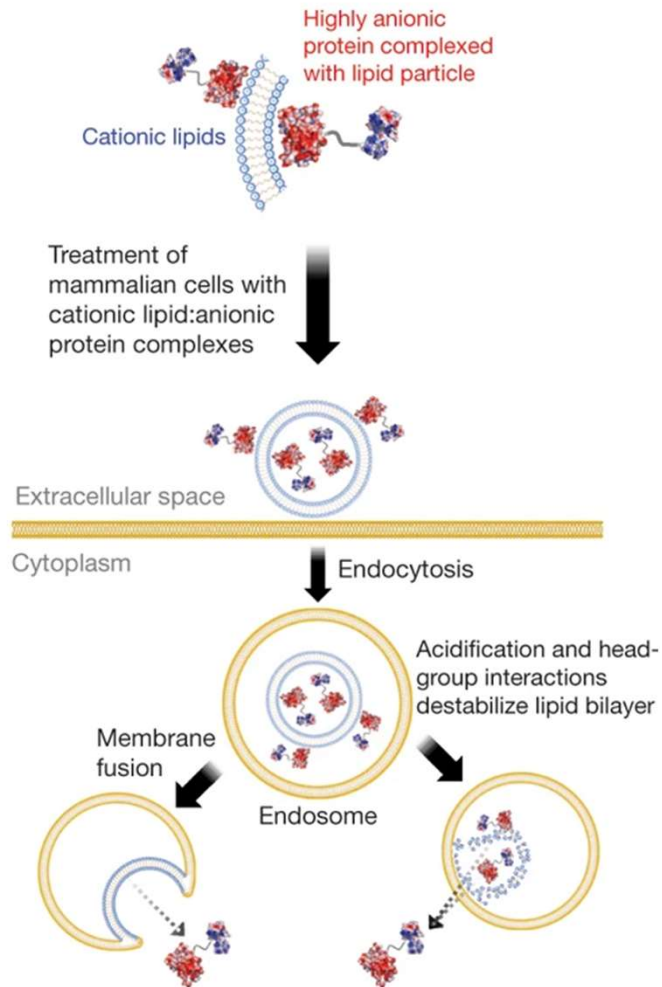
- Interaction with membranes
- Endocytosis
- Escape from endosomes

⌘ All of them are cytotoxic.



Methods of delivering CRISPR/Cas9 RNP ①: Synthetic lipid nanoparticles

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- the Cas9/sgRNA RNP is negatively charged
→ Encapsulated by lipid molecules in liposomes.
- Most commonly used in research applications.

Ex) lipofectamine CRISPRMAX
(Commercially available reagents dedicated to CRISPR transfection, the genome editing efficiencies achieved 55%, 75% and 85% in human iPSCs, mouse ES cells and HEK293FT cells)

- HDR achieved

Zuris, J., Thompson, D., Shu, Y. *et al. Nat Biotechnol* **33**, 73–80 (2015).

Methods of delivering CRISPR/Cas9 RNP ② : polymers

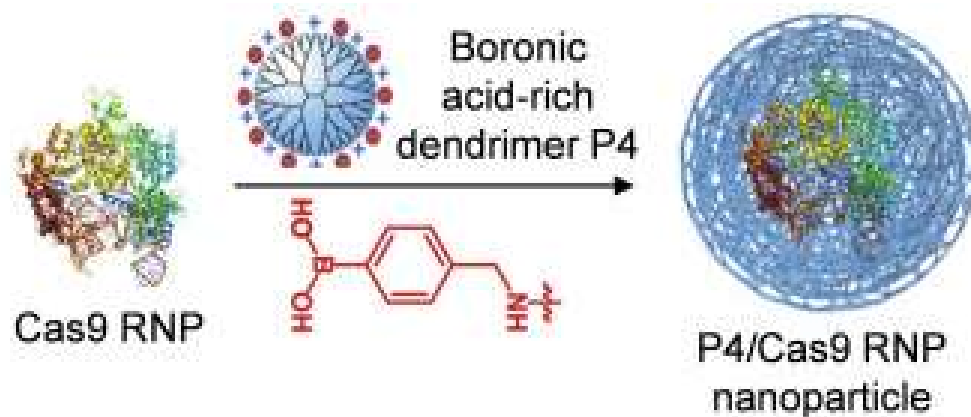
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- Form complexes with Cas9 RNPs using polymers

Advantages

- Easy to synthesize
- Flexible structure and composition
- Easy to functionalize
- Degradable

Ex) PBA-rich dendrimer

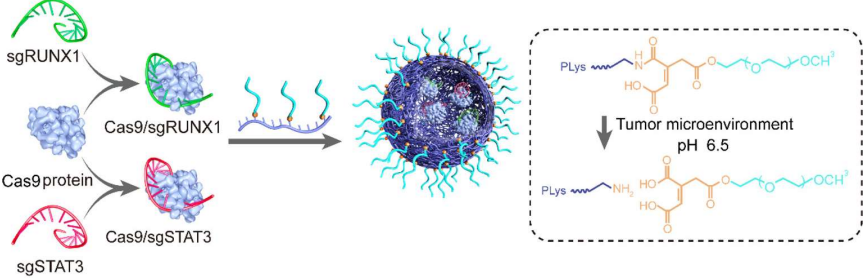


Interaction of phenylboronic acid (PBA) with proteins to form a more stable complex

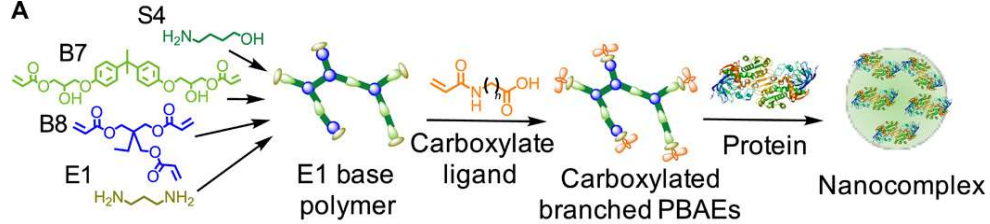
- 20~40% in del (293T-EGFP cells)

Methods of delivering CRISPR/Cas9 RNP ② : polymers

PEGylated PLL

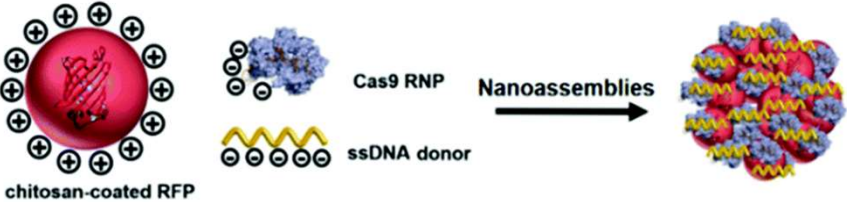


PBAE (poly(β -amino ester) nanoparticles)



• HDR achieved

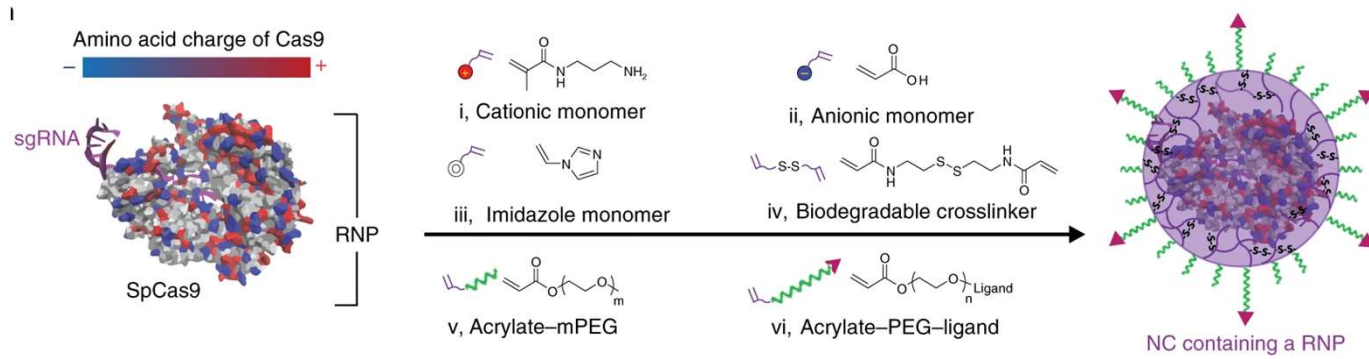
Chitosan (CS) nanoparticles



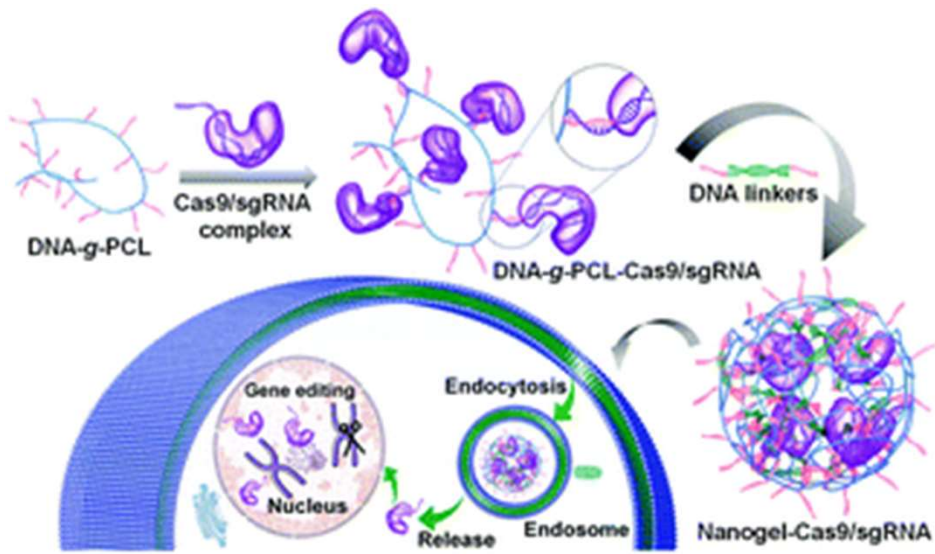
• HDR achieved

Methods of delivering CRISPR/Cas9 RNP ③ :nanogel

nano capsule



G. Chen, A.A. Abdeen, et al. *Nat. Nanotechnol.*, 14 (2019), pp. 974-980



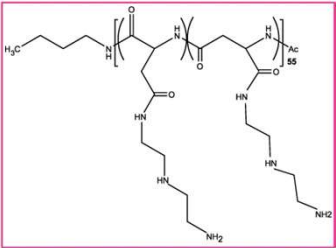
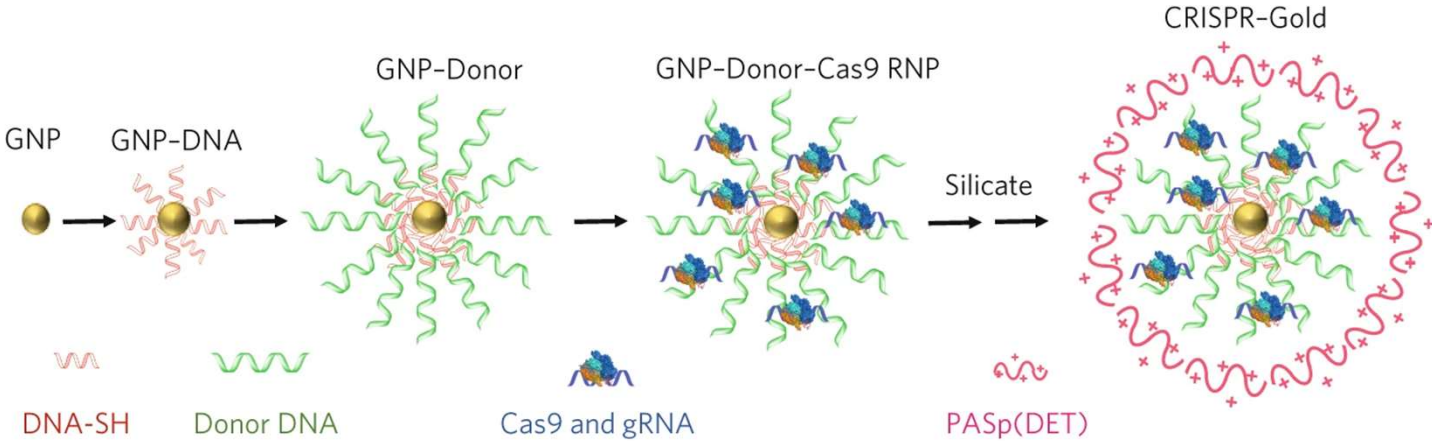
• 18.7% in del

Zhang S, Shen J, Li D, Cheng Y. *Theranostics* 2021; 11(2):614-648. ¹⁷

Methods of delivering CRISPR/Cas9 RNP ④ : Inorganic nanoparticles

- Use inorganic particles, such as metals, to form the complex.

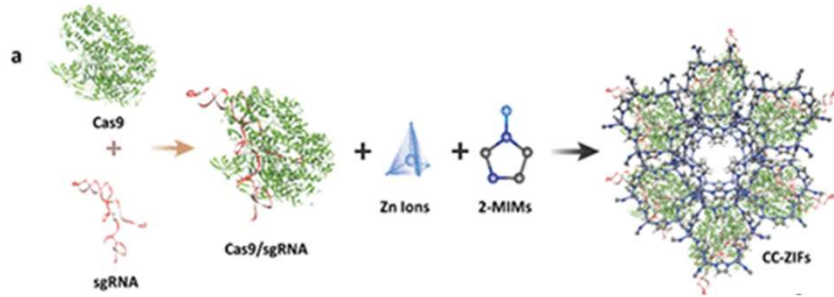
AuNP



• HDR achieved (~5%)

Methods of delivering CRISPR/Cas9 RNP ④ : Inorganic nanoparticles

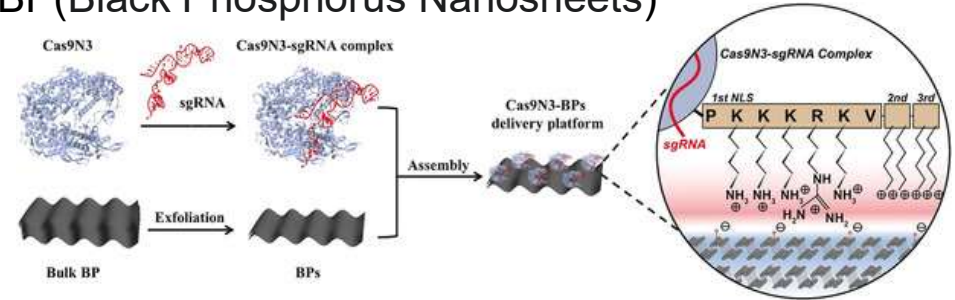
MOF(Metal-organic frameworks)



- 30~60% in del
- HDR achieved

Shahad K. et al.
Journal of the American Chemical Society **2018** 140 (1), 143-146

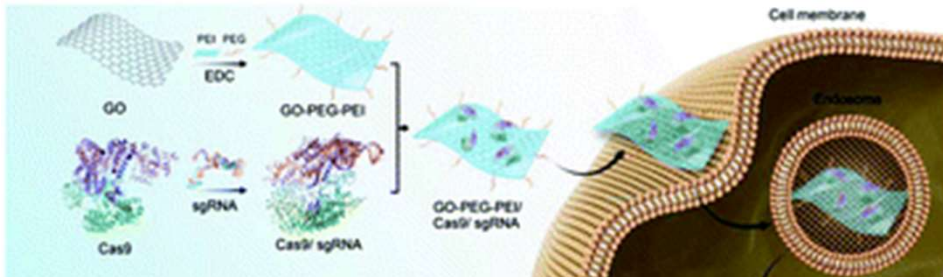
BP(Black Phosphorus Nanosheets)



- 17~32% in del

W. Zhou, H. Cui, L. Ying, X.-F. Yu, *Angew. Chem. Int. Ed.* **2018**, 57, 10268.

GO(graphene oxide)



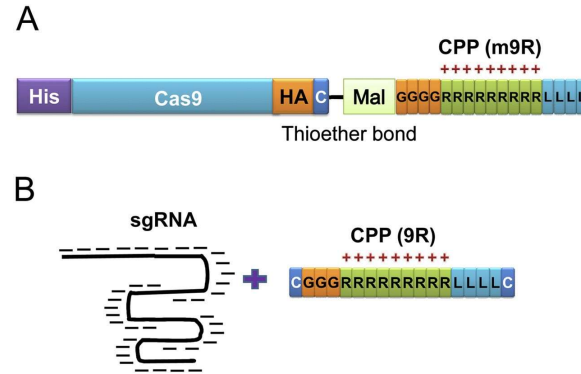
- 30~60% in del

Yue, H., Zhou, X., Cheng, M., & Xing, D. (2018). *Nanoscale*, 10(3), 1063–1071.

Zhang S, Shen J, Li D, Cheng Y. *Theranostics* 2021; 11(2):614-648. 19

Methods of delivering CRISPR/Cas9 RNP ⑤ : CPP conjugation

ex : Cas9 +CPP / CPP · sgRNA complex

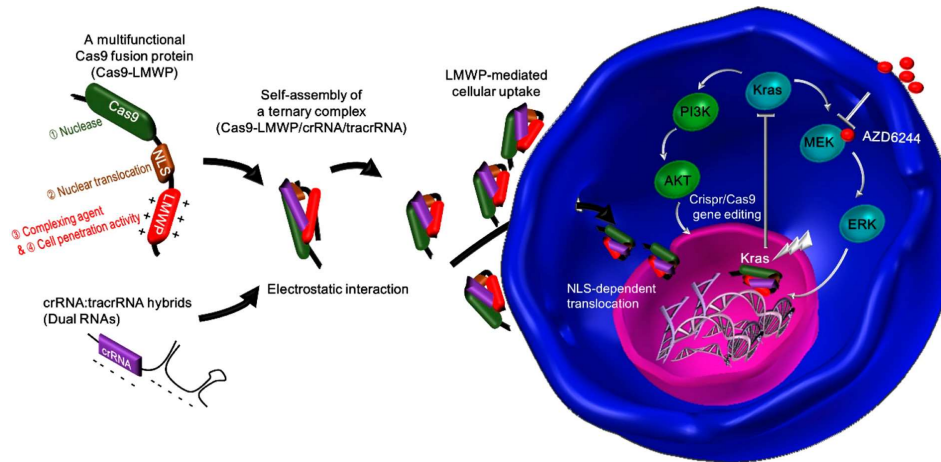


Cas9-Smal-CPP
 +sgRNA-CPP complex

16% in del (HEK293T *in vitro*)

Ramakrishna, S., Kwaku Dad, A. B., et al. *Genome Research*, 24(6), 1020–1027.

ex : Cas9-LMWP-sgRNA ternary Cas9 RNP



Cas9-NLS-LMWP(CPP) protein
 +sgRNA

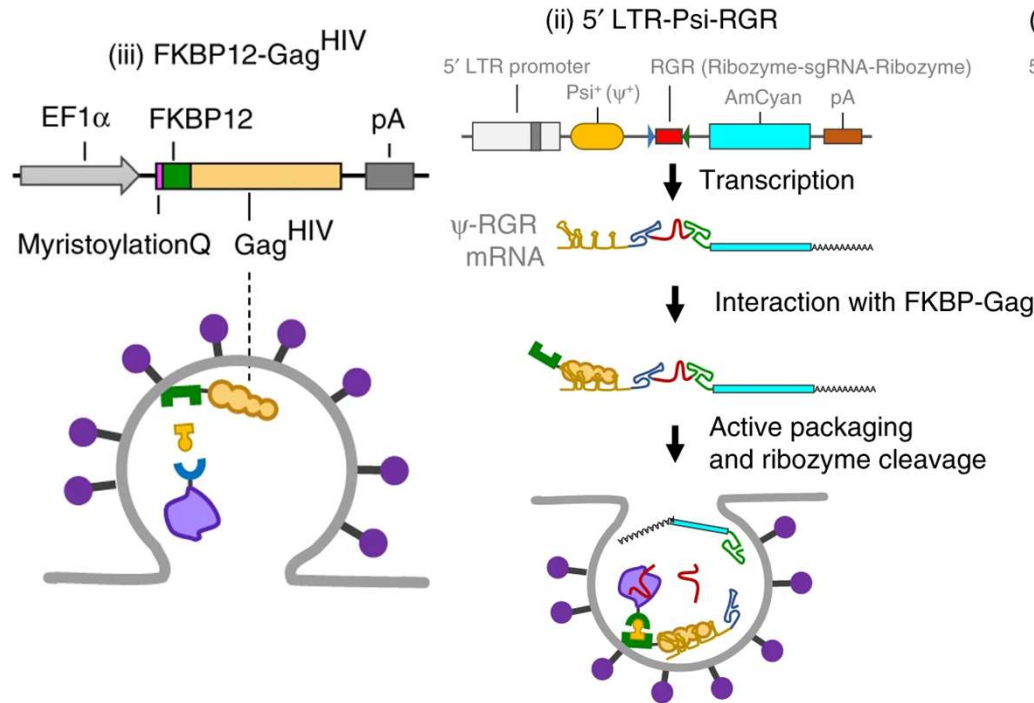
43.9% in del (A549 cells *in vitro*)

Seung Min Kim, et al. *ACS Nano* 2018 12 (8), 7750-7760

Methods of delivering CRISPR/Cas9 RNP ⑥ : Biological Production

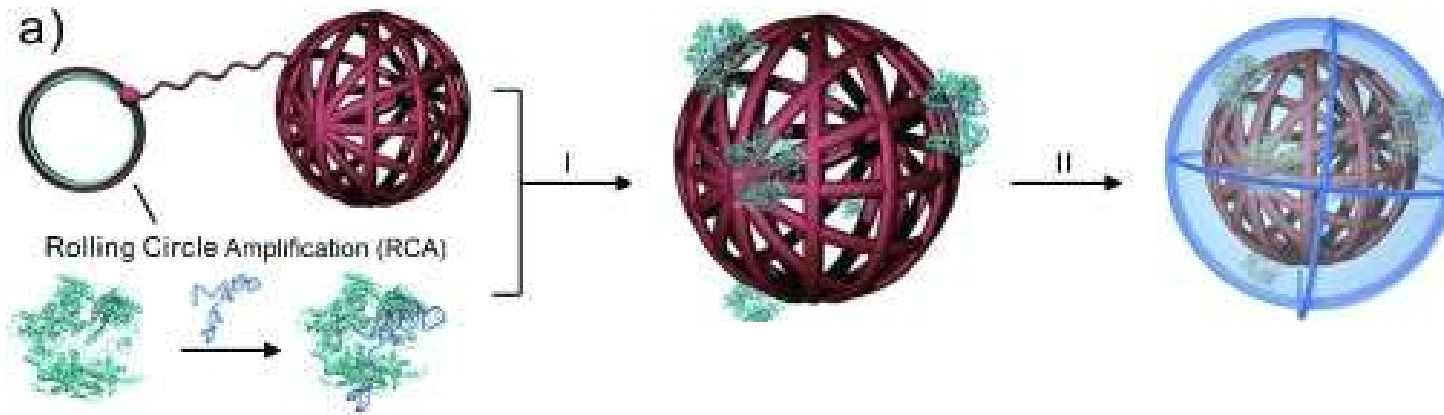
- Viral vectors, extracellular vesicles, etc.

Ex) NanoMEDIC



- Expression of three viral proteins (VSV, HIV-Gag, and LM) to form virus-like vesicles
- Cas9 protein is incorporated into EVs through interaction between FRB and FKBP
- sgRNA is incorporated into EVs by interaction between GAG and ψ⁺.
- Introduced into target cells by viral proteins on budding extracellular vesicles
- up to 92% exon skipping (DMD patient iPSCs)

- DNA nanoclew



- Various delivery methods for CRISPR-Cas9 RNPs have been developed.
- Many of them form nanoparticles or covalent complexes that are permeable to cell membranes and are transferred into cells via endocytosis.
- Nanoparticle materials are diverse(Lipids, polymers, inorganic particles, peptides, etc.)

Most of the existing systems have focused on gene deletion by NHEJ, and few have achieved gene transfer by HDR.

- HDR achieved
 - liposome : 5~85% in del/ 8~16% HDR
 - AuNP : 14.6~34% in del / ~5% HDR
 - MOF : 30~60% in del / 20~% HDR
 - Chitosan (CS) nanoparticles : 16.9~55.8% in del /12.5 % HDR
 - PBAE(poly(β -amino ester) nanoparticles) : 47~77 % in del / 4% HDR

HDR is difficult to achieve with existing systems

- Low efficiency compared to NHEJ → Cytotoxicity of carriers (especially cationic carrier) occurs when trying to increase the concentration
- Co-localization of donor DNA is required(NHEJ : only RNP is required)

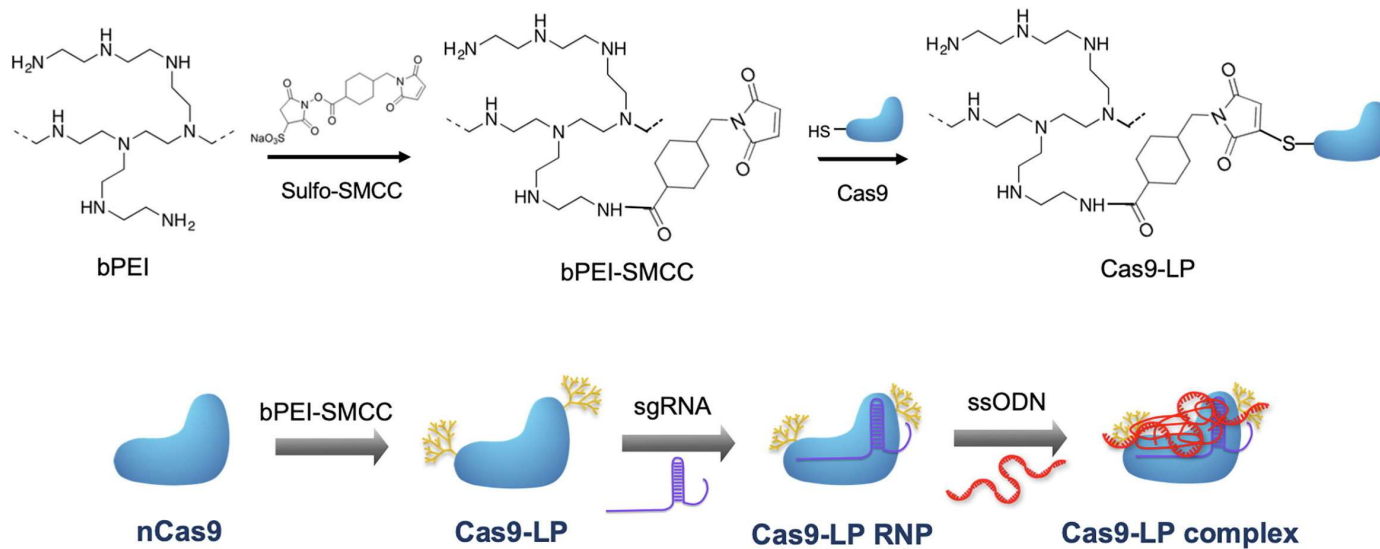
Zhang S, Shen J, Li D, Cheng Y. *Theranostics* 2021; 11(2):614-648.

Yoo Kyung Kang, et al.

Journal of Industrial and Engineering Chemistry, Volume 102, 2021, Pages 241-250,

Challenge to create a system that can generate HDR with high yield

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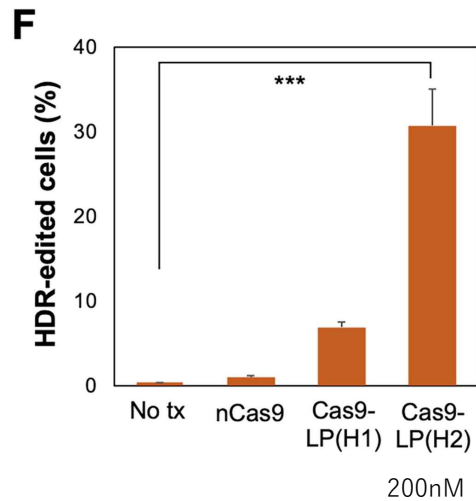
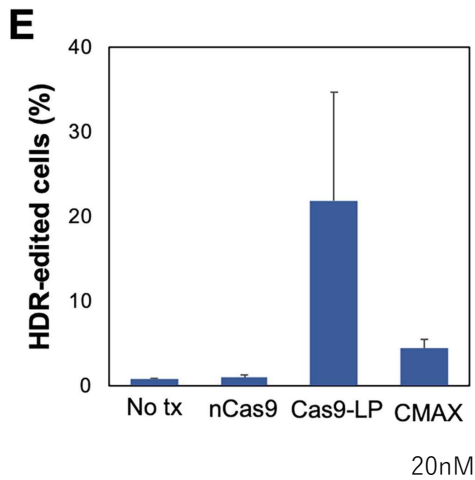
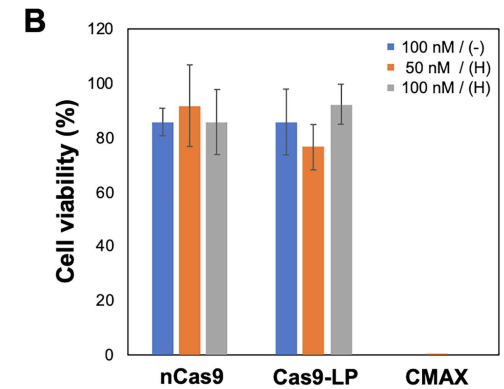
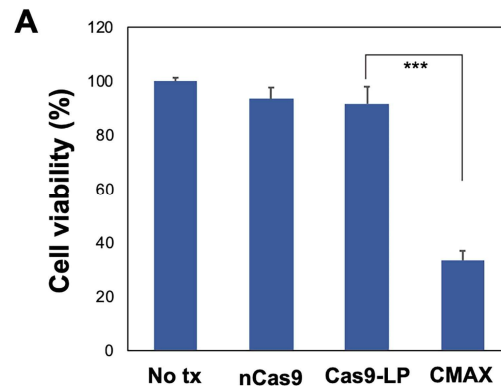
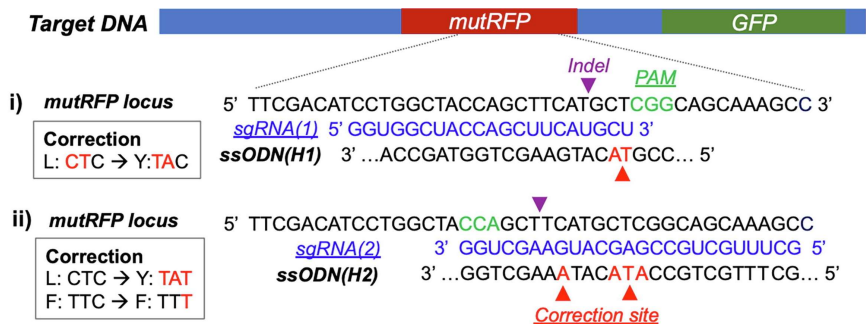


- As few carriers as possible → Aim to reduce toxicity
- Interaction and complexation with ssODN by charge
→ Enables co-localization with donor DNA

Yoo Kyung Kang, et al.

Journal of Industrial and Engineering Chemistry, Volume 102, 2021, Pages 241-250, 25

Challenge to create a system that can generate HDR with high yield



- Low cytotoxicity
- Higher concentration could be achieved
- HDR : up to 31% (exceeding CRISPRMAX)
- Still limited to use in vitro → awaiting vivo application

- ① Introduction
- ② Methods of delivering CRISPR/Cas9 RNP
- ③ Delivery of systems that cause HDR with high yields
- ④ Summary

introduction

- CRISPR-Cas9 is attracting attention as a powerful tool for genome editing therapy.
- Drug discovery using Cas9 has already begun, and some have advanced to clinical trials.
- Delivery of Cas9 in the form of RNPs has many advantages over other methods, and delivery methods are being developed to achieve these advantages.

Methods of delivering CRISPR/Cas9 RNP

- Intracellular delivery of CRISPR-Cas9 RNPs is generally performed by linking them with membrane-permeable materials.
- The materials of the complexes can be lipids, polymers, inorganic particles, peptides, etc.
- Most of the currently developed delivery methods target only deletion editing, and only a limited number of systems are capable of HDR-mediated gene transfer, but the development of delivery methods that enable this is underway.