

Artificial Enzyme

-Computational Design of Catalytic Functions-

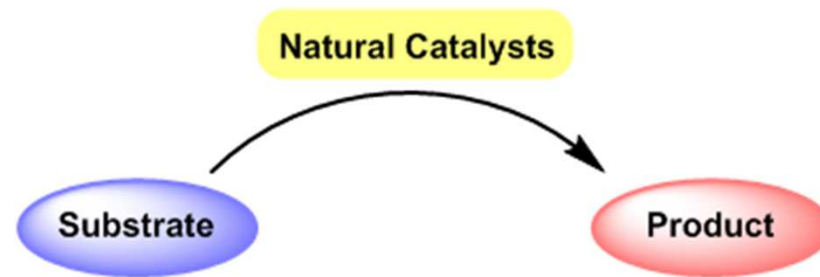
Literature Seminar

2014.2.3 (Mon.)

Yusuke Shimizu (B4)

Biocatalysis

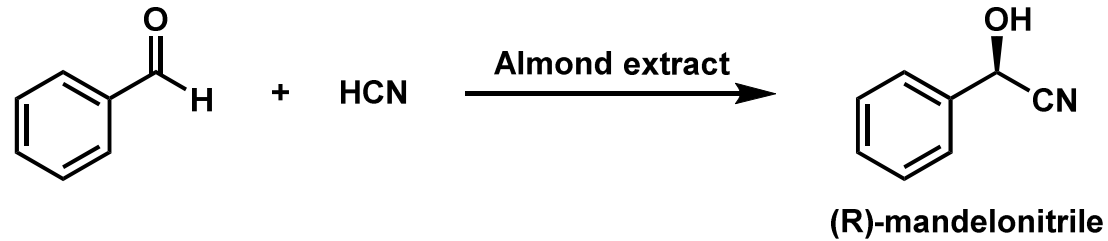
Biocatalysis = Application of enzymes and microbes to chemical transformations



Humans have utilized biocatalysis in the fermentation processes for millennia.

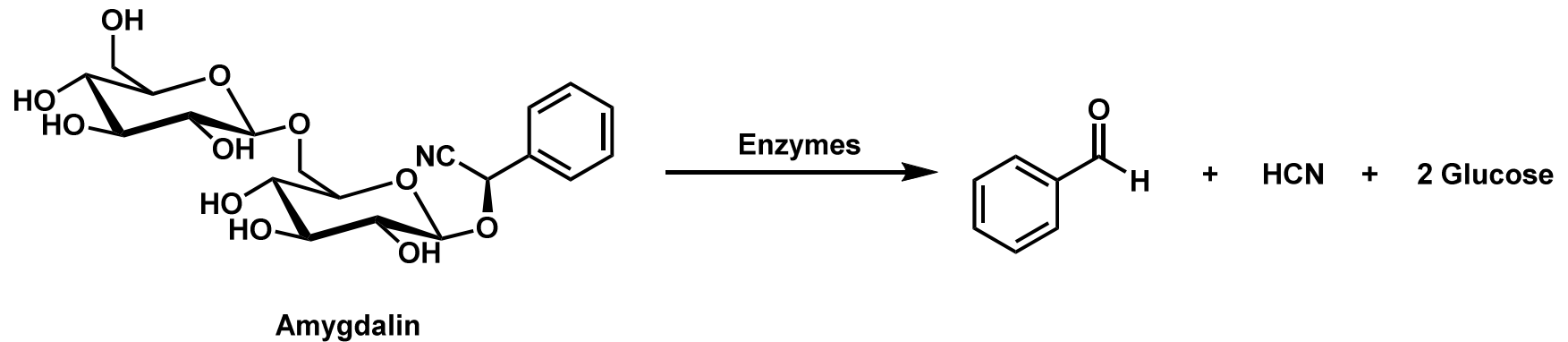


Biocatalysis



Rosenthaler, L. *Biochem. Z.* **1908**, 14, 238

Actual “biocatalyst” is an enzyme related to cyanogenesis



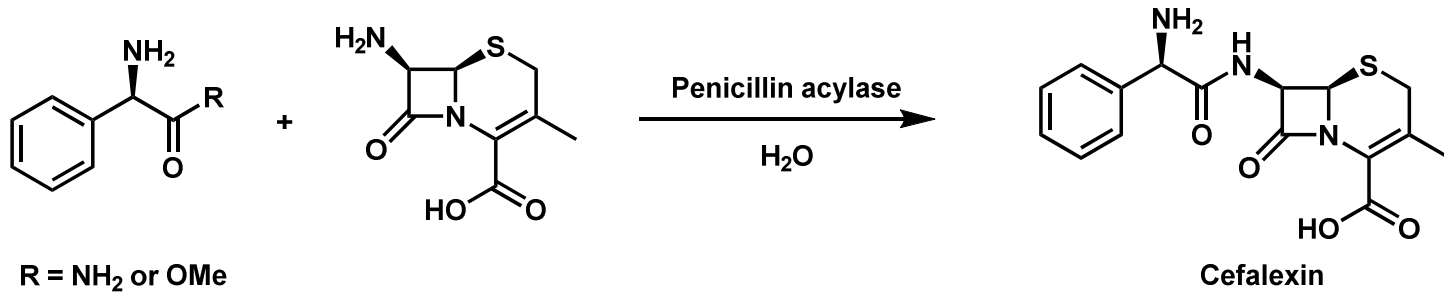
What is Enzyme?

- First isolation in 1833 by Payen. He discovered and isolated amylase.
- Kühne coined term *enzyme* means “in yeast” in 1876
- Sumner crystallized urease, and showed enzymes are proteins in 1926.
- The first crystal structure was obtained in 1965 via X-ray crystallography

Enzyme is a protein catalyst , which is...

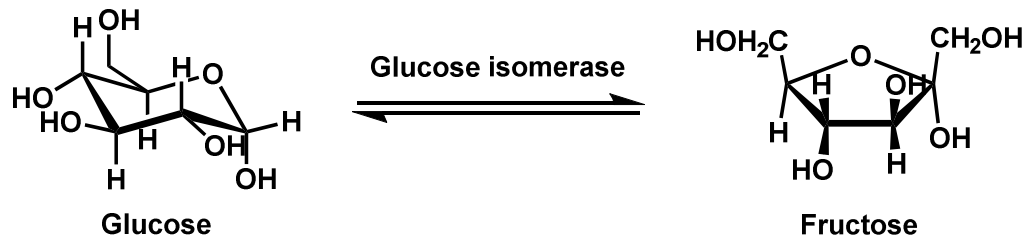
- Highly selective (chemo-, regio-, diastereo-, and enantio-selective)
- Activated in mild condition (typically pH 5–8 and 20–40°C)
- Environment friendly (completely degraded in the environment)
- Generally very efficient

Enzymes in Industry



Bruggink, A. *Chimia*, 1996, 50, 431

Semi-synthetic antibiotics



Jensen, V.J., Rough, S. *Methods Enzymol.* 1987, 136, 356

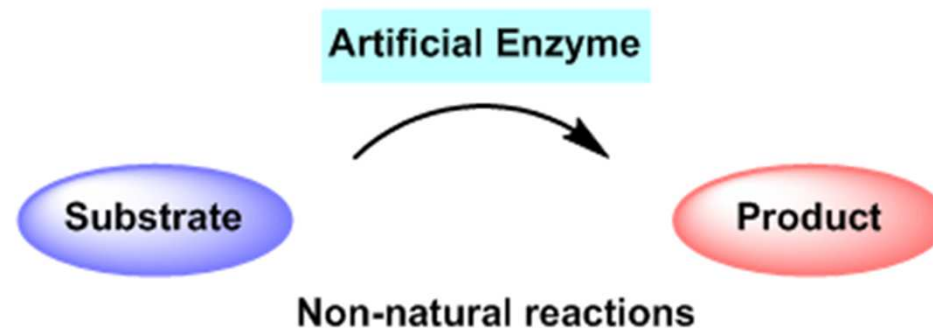
Convert glucose to the sweeter-tasting fructose



Laundry-detergent contain enzymes

Artificial Enzyme

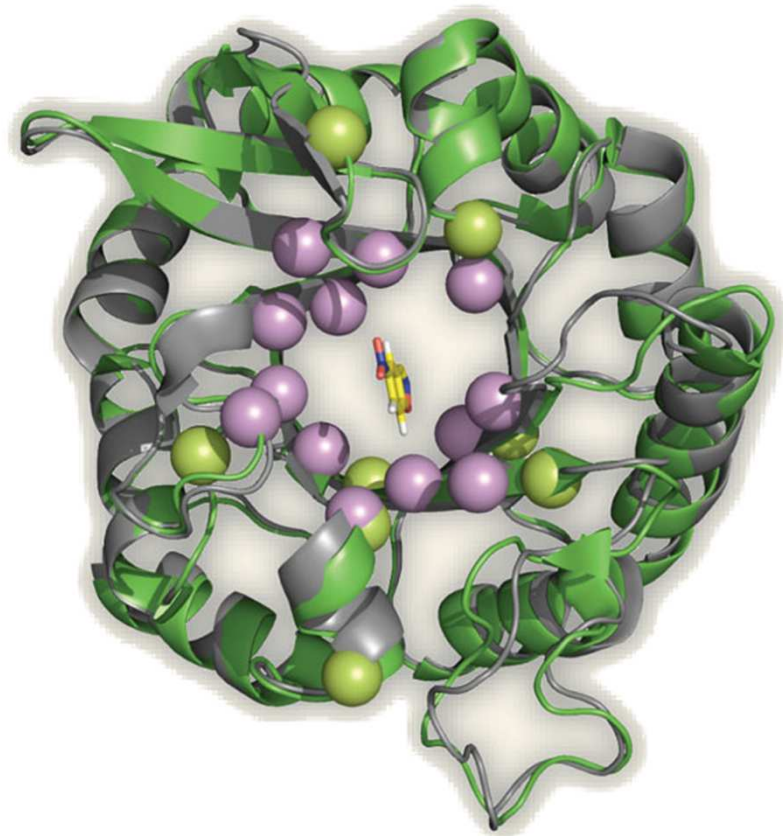
Protein engineering technologies has developed to design and synthesize molecules with the attributes of enzymes (selective, proficient, green, nontoxic, and biodegradable) for non-natural reactions



Today, I introduce development of approach to enzyme design and finally, *de novo* Computational Enzyme Design.

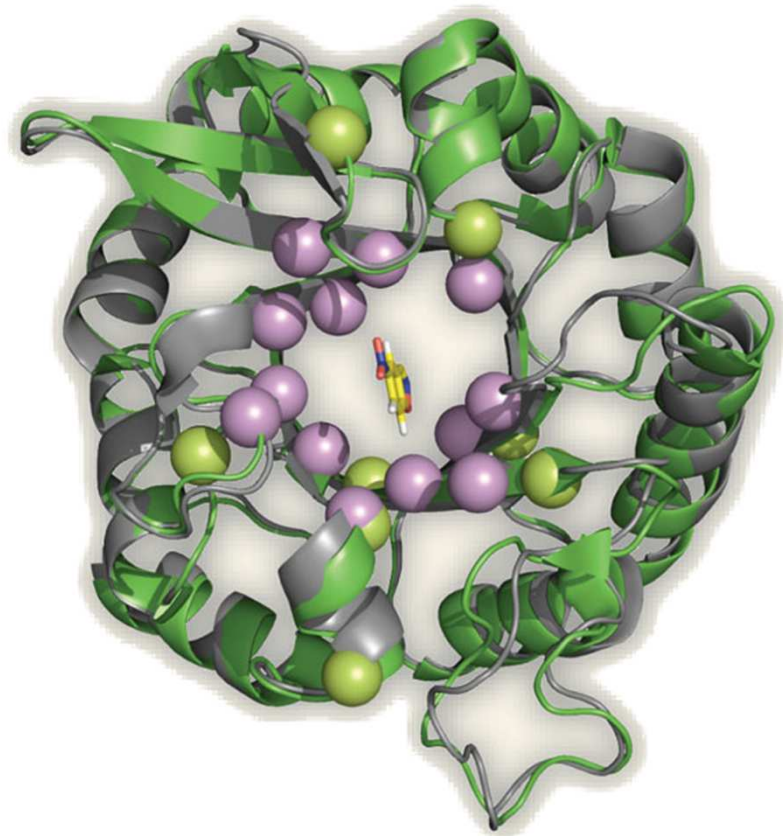
Contents

1. Introduction
2. Catalytic Antibodies
3. Directed Evolution
4. Computational Enzyme Design
5. Summary & Future Outlook



Contents

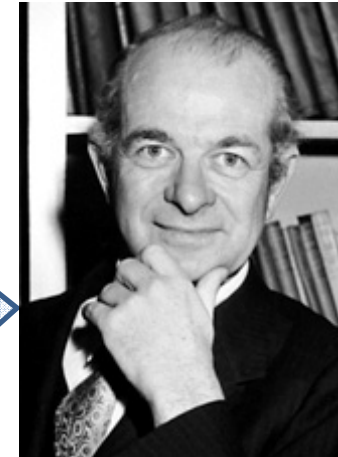
1. Introduction
2. Catalytic Antibodies
3. Directed Evolution
4. Computational Enzyme Design
5. Summary & Future Outlook



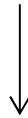
Pauling's hypothesis

I think that enzymes are molecules that are complementary in structure to the activated complexes of the reaction that they catalyze, that is, to the molecular configuration that is intermediate between reacting substances and the products of reaction for these catalyzed process

Pauling, L. *Nature*, 1948, 161, 707



Linus Pauling



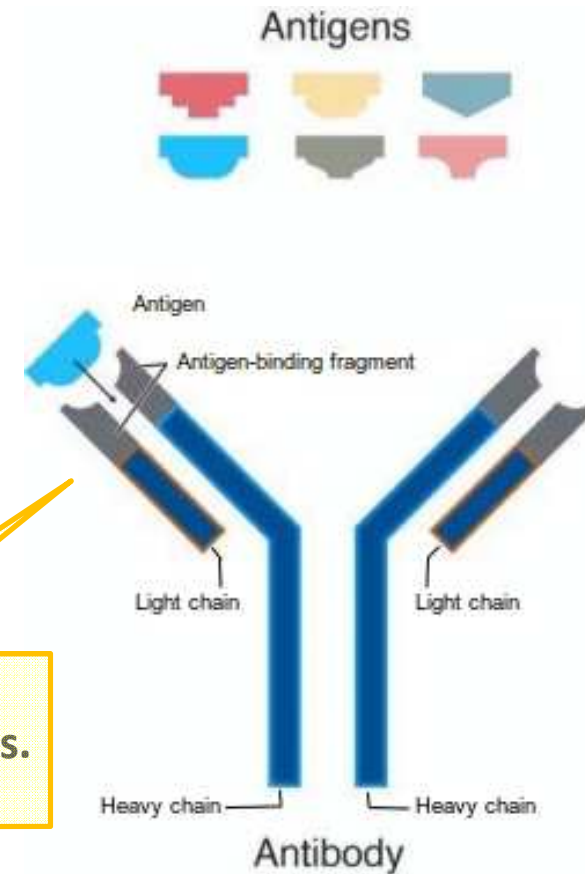
The transition state analog of a particular enzymatic reaction would be bound tightly to enzymes. (Concept of enzyme inhibitors)

Catalytic Antibodies

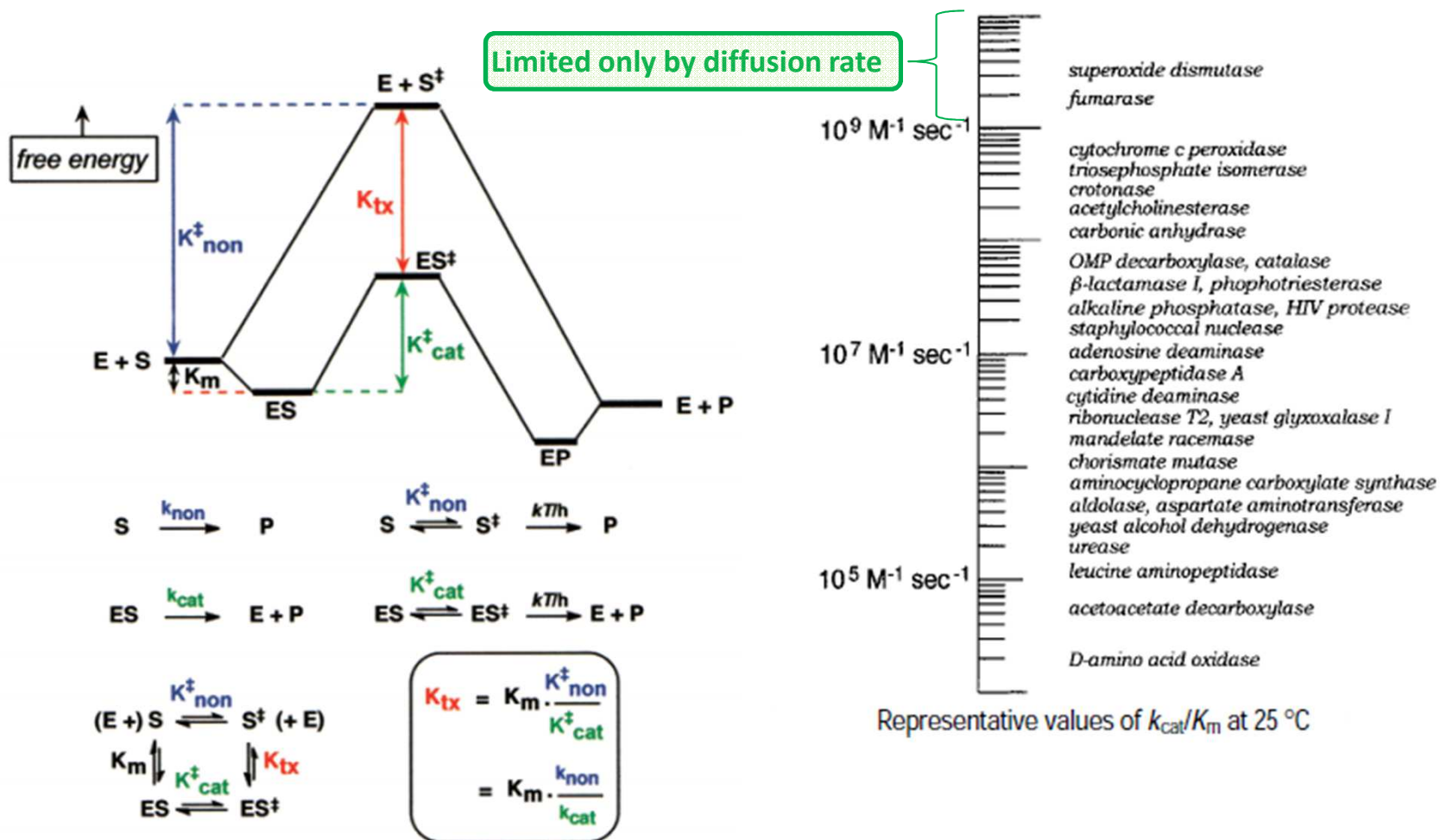
According to Pauling's hypothesis, conversely, a receptor designed to optimally bind a suitable analog of a transition state would achieve the catalytic function of an enzyme.

The immunological reservoir provides the variety of receptor sites with the required specificities for such a study. The combining sites of antibodies have been considered as useful templates for simulating the environment of an enzyme active site

Antibodies specifically bind to corresponding antigens.



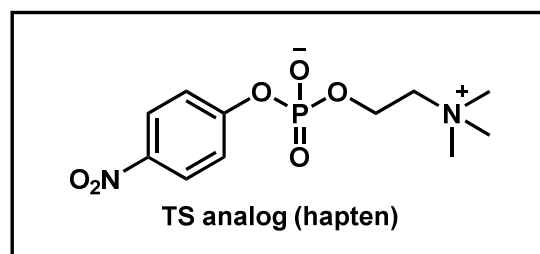
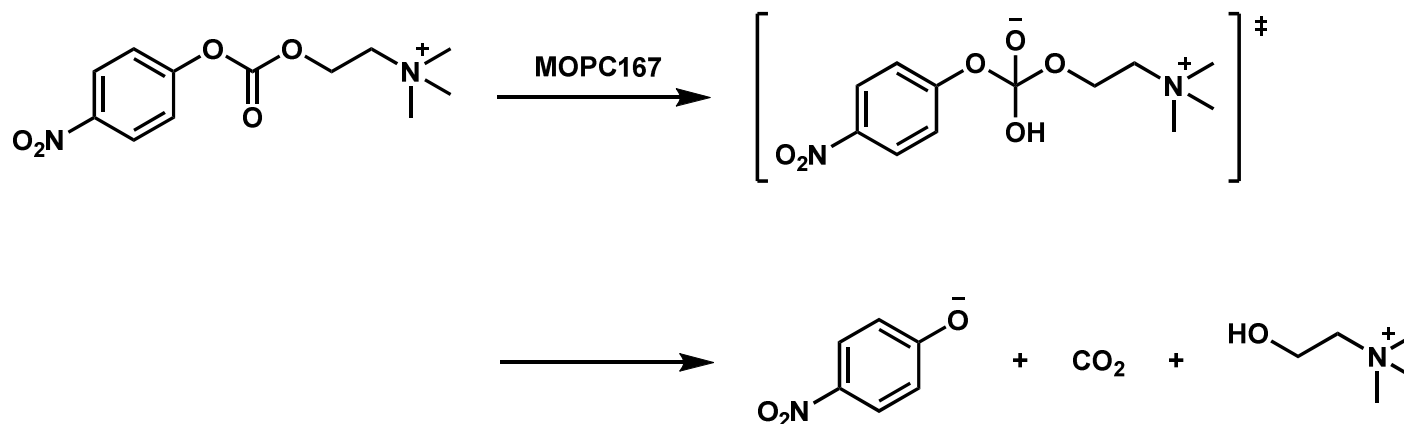
Kinetics



Wolfenden, R., Snider M. J. *Acc. Chem. Res.* **2001**, 34, 938

Pioneering work by Schulz

Pioneering work of catalytic antibody using IgA MOPC167

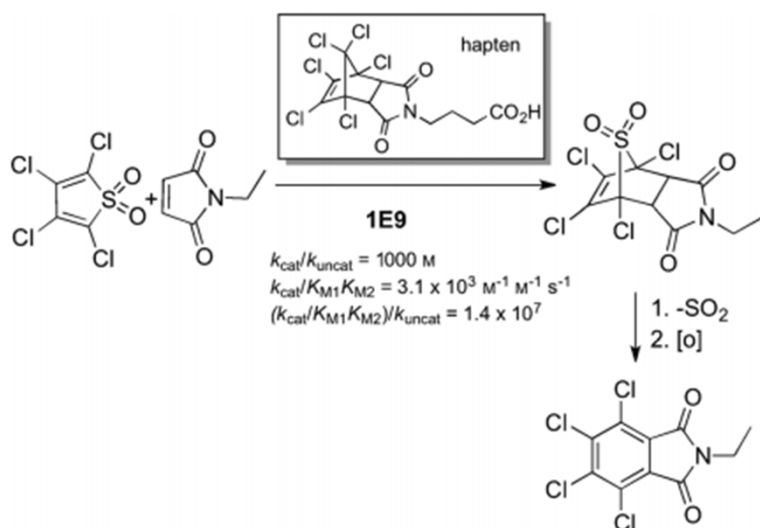


$$\begin{aligned}K_M &= 208 \mu\text{M} \\K_{\text{cat}} &= 6.0 \cdot 10^{-3} \text{ sec}^{-1} \\k_{\text{cat}}/K_M &= 28.8 \text{ M}^{-1}\text{s}^{-1} \\k_{\text{cat}}/k_{\text{uncat}} &= 770\end{aligned}$$

Schulz, P.G., *et al. Science*, **1986**, 234, 1570

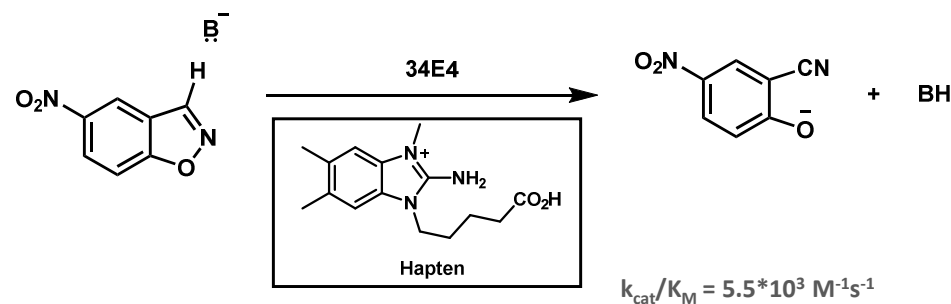
Achievement

A number of catalytic antibodies catalyze natural and non-natural reaction were generated.



Diels-Alder reaction

Hilvert, D., et al. *J. Am. Chem. Soc.* **1989**, 111, 9262



Kemp elimination

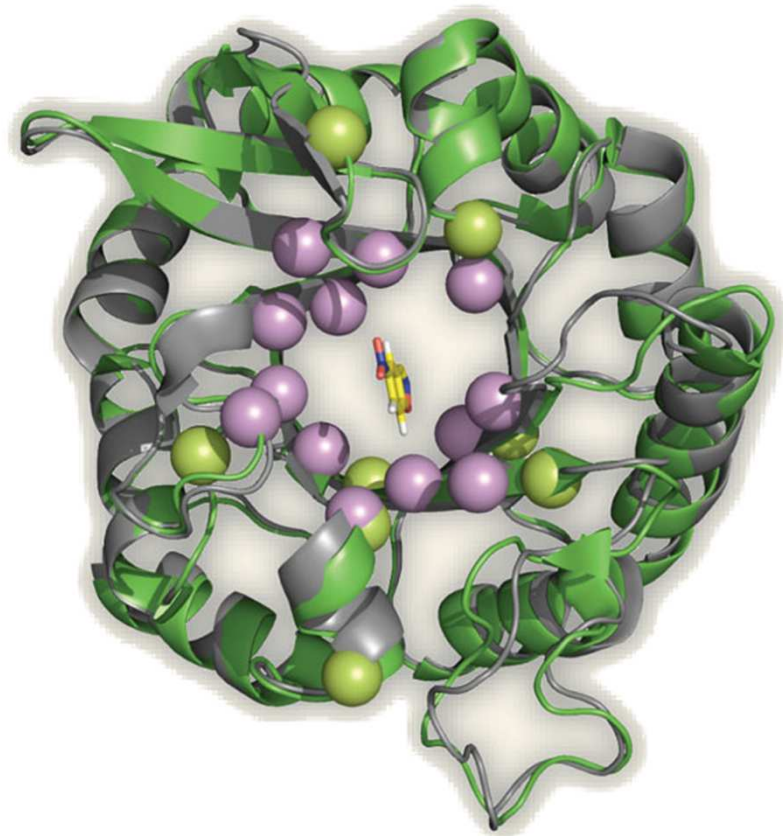
Hilvert, D., et al. *Nature*, **1995**, 373, 228

Limitations

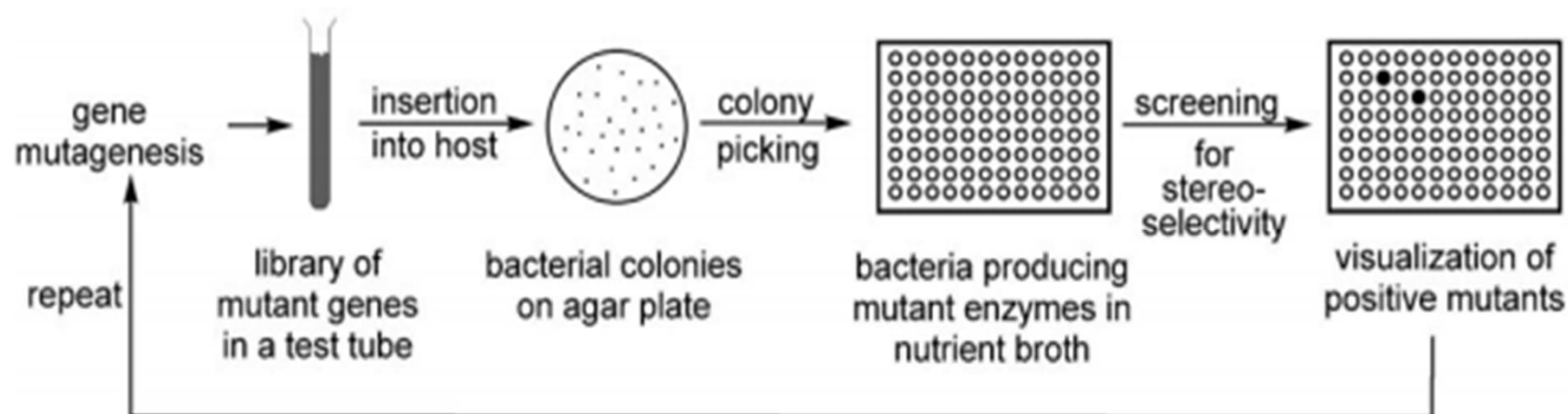
- Lower binding constants
- Lack of covalent binding and catalysis
- Smaller buried surface area
- Inadequacies of the immunoglobulin fold
- Product inhibition
- The comparatively low stability of the immunoglobulin fold
- High cost of producing antibody catalysts

Contents

1. Introduction
2. Catalytic Antibodies
3. Directed Evolution
4. Computational Enzyme Design
5. Summary & Future Outlook



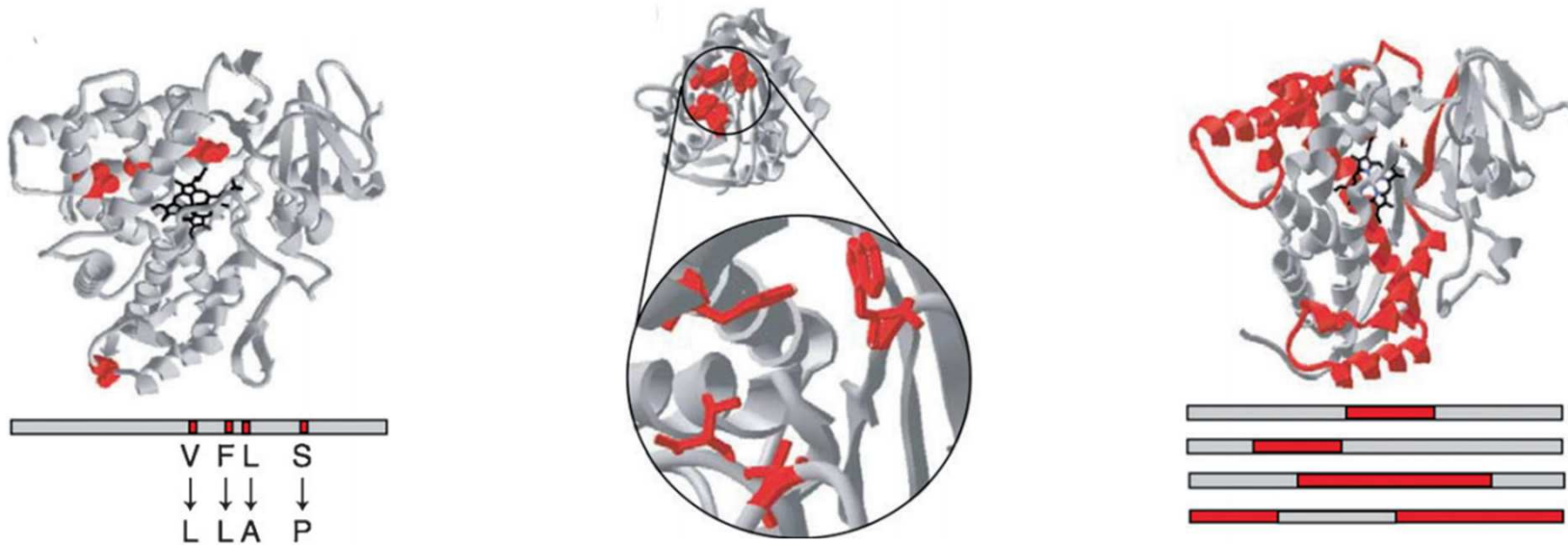
Directed Evolution



Directed evolution is a molecular biology methods to modify biocatalysts via *in vitro* version of “Darwinian evolution”

Directed evolution provide improved enzymatic activity, thermostability, tolerance to organic solvent, substrate specificity, enantioselectivity and so on.

Mutagenesis



Random mutagenesis

Site saturation mutagenesis

Sequence recombination

Random mutagenesis : No structural or mechanistic information is required

Site saturation mutagenesis : Require prior structural or biochemical knowledge

Dramatic functional alteration can be sought

Sequence recombination : Diversity can be further expanded

Challenges

- Identification of variants that have desired improvements out of large set of sequences. (e.g. two mutations anywhere in 200 amino-acid protein have 7,183,900 possibilities)
- Set up of high-throughput assays which can detect slight improvements

Application – Sitagliptin Manufacture

Route A

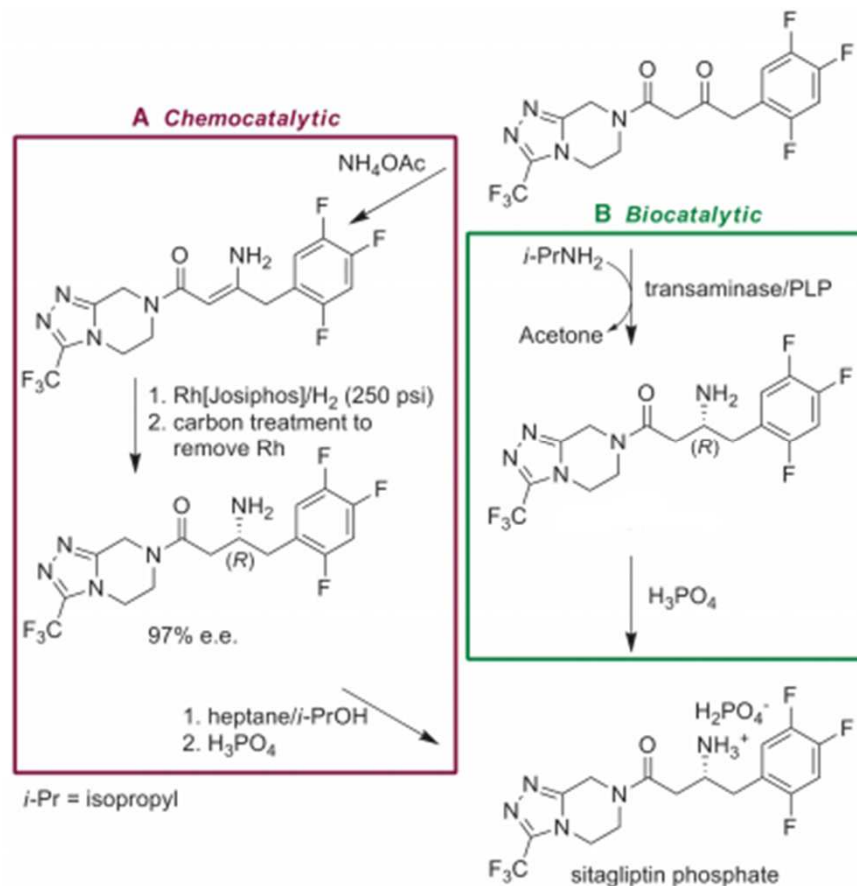
- Need for high pressure H₂
- Use and removal of precious and toxic Rh
- Ligand screening and synthesis
- Insufficient stereoselectivity

Route B

- Limited substrate range
- Low turn over numbers
- Stability to chemical processes conditions



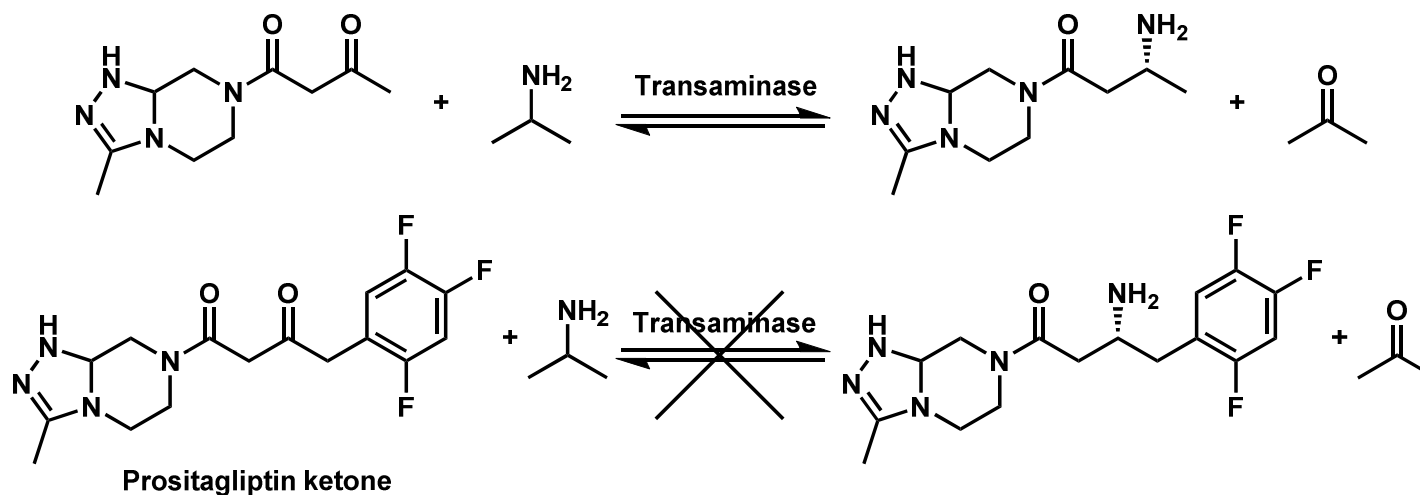
Directed Evolution



Savile, C. K., et al. *Science*, 2010, 329, 305

Substrate range of transaminase

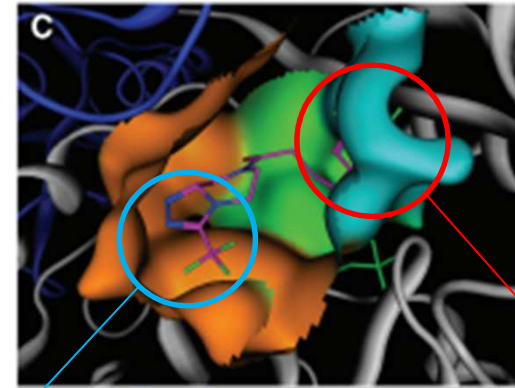
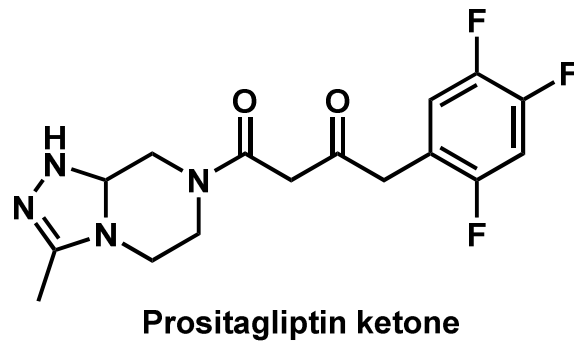
Transaminases have a limited substrate range;
Most of them accept only substrates with a substituent
no longer than methyl group at the position adjacent to the ketone



Screening a variety of transaminases provided no detectable activity.

Docking Study

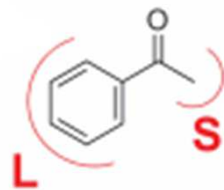
Using (R)-selective transaminase ATA-117, homology model was generated.



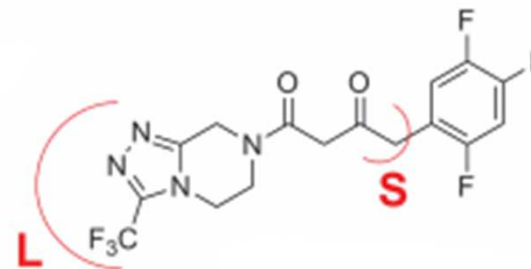
Undesired interactions

Steric interference

Prositagliptin ketone has problems in binding both with **small pocket** and **large pocket**.



substrate of ATA-117

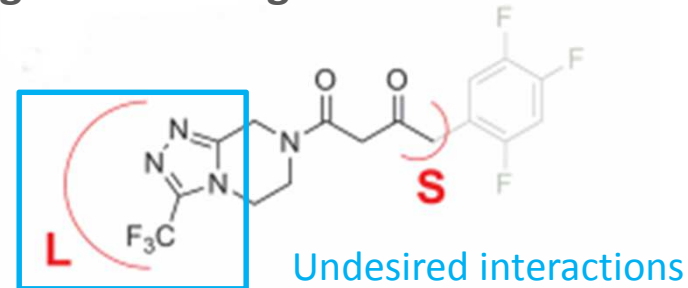


Savile, C. K., et al. *Science*, 2010, 329, 305

Round 1

Firstly, the large binding pocket was engineered using truncated substrate

Consistent with the model,
this substrate was poorly active.
(4% conversion of 2g/L substrate)



Site saturation mutagenesis around the large pocket.

The best variant containing S²²³→P²²³ showed 11-fold activity

A small library for potential activity on prositagliptin ketone

Analysis of the model suggested four residues (V⁶⁹, F¹²², T²⁸³, and A²⁸⁴)
potentially interact with trifluorophenyl group. Savile, C. K., et al. *Science*, 2010, 329, 305

Round 1

ATA-117: S²²³→P²²³(S223P) variant

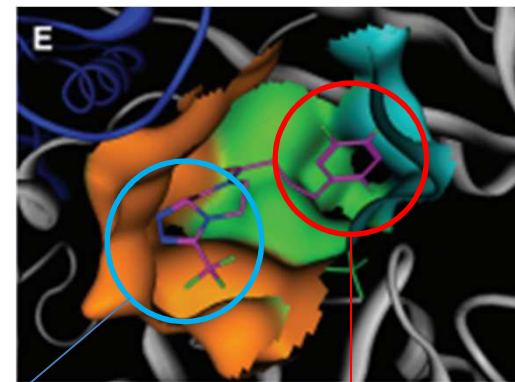


Saturation mutagenesis on V⁶⁹, F¹²², T²⁸³, and A²⁸⁴ individually & Combinatorial library based on structural considerations. (V69GA, F122AVLIG, T283GAS, and A284GF; 216 variants)

Single site saturation library provided no active variant.
Y26H, V65A, V69G, F122I, A284G provided initial active variant.
(0.7% conversion of 2g/L ketone)



Round 2



Relieved steric interference

Effective interaction Savile, C. K., *et al. Science*, 2010, 329, 305

Round 2

Initial active variant



Library screening

Variant containing 12 mutations showed 75-fold activity

Despite these enhancement, this catalyst was not yet practical...

- Organic co-solvent (low water-solubility of ketone)
- Higher temperature (rate and solubility enhancement)
- Large excess of $i\text{PrNH}_2$ (transamination is equilibrium controlled)

Transaminase have to withstand these harsh conditions.



Further evolution

Round 3-11

Transaminase with enhanced activity



Library generated by a variety of methods (total 36480 variants)
Rendering condition more stringent with the rising tolerance

Final screening conditions

Substrate : 2 → 100g/L

iPrNH₂ : 0.5 → 1M

Co-solvent : 5 → 50% DMSO

pH : 7.5 to 8.5

Temp. : 22°C to 45°C

Process tolerant transaminase containing 27 mutations

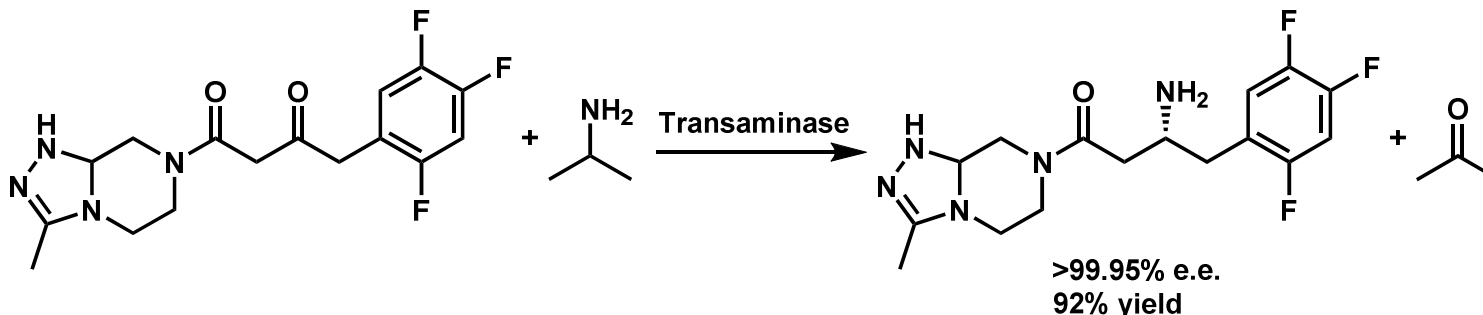
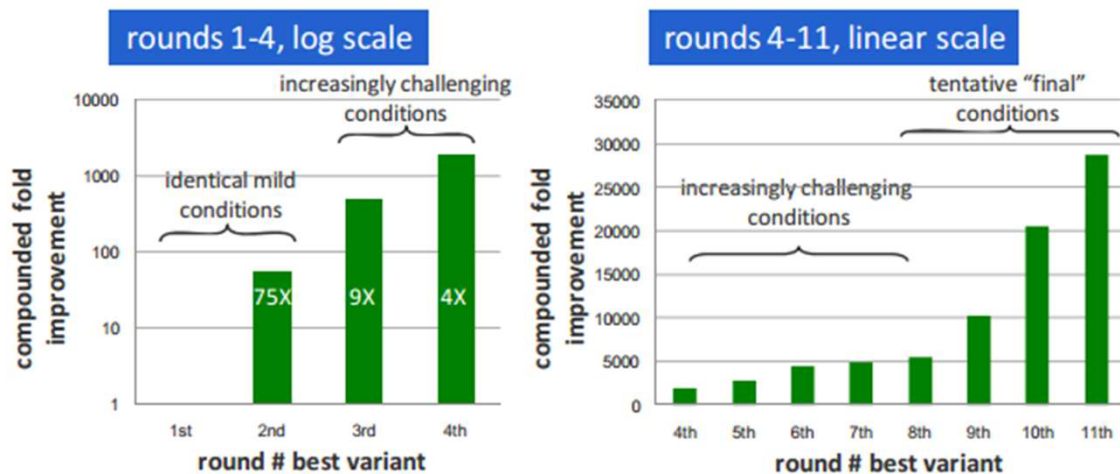
§3 Directed Evolution

Summary of evolution

Substrate	Added Mutations [*]	[Substrate] in g/l	Assay changes	Round identified	Improvement over parent [†]
1	ATA-117	2		–	N/A
1	G136Y	2		1a	6
1	S223P	2		1a	11
2	S223P	2		1a	Not active
2	Y26H; [‡] V65A; [‡] V69G; F122I; A284G	2		1b	first active
2	H62T; G136Y; E137I; V199I; A209L; T282S	2		2	75
2	S8P; H26Y; G69C; M94I; I137T; G215C	5	5% DMSO to 5% MeOH; RT to 30°C	3	9
2	L61Y; C69T; Y136F; T137E	10	0.5 to 1 M iPrNH ₂ ; pH 7.5 to pH 8.5	4	4
2	D81G; I94L; I96L; T178S; L269P; P297S; S321P	40	5 to 10% MeOH; 30 to 45°C	5	1.4
2	Y60F; L94I; A169L; S178T; G217N; L273Y	100	10 to 20% MeOH	6	1.6
2	S124H	100	20% MeOH to 25% DMSO	7	1.1
2	I122M; H124N	100		8	1.1
2	Q329H	100		9	1.9
2	N124T; Y150S; V152C; H329Q	50	25 to 50% DMSO; 0.5% acetone	10	2
2	S126T	50		11	1.4

Savile, C. K., *et al. Science*, **2010**, 329, 305

Improvements

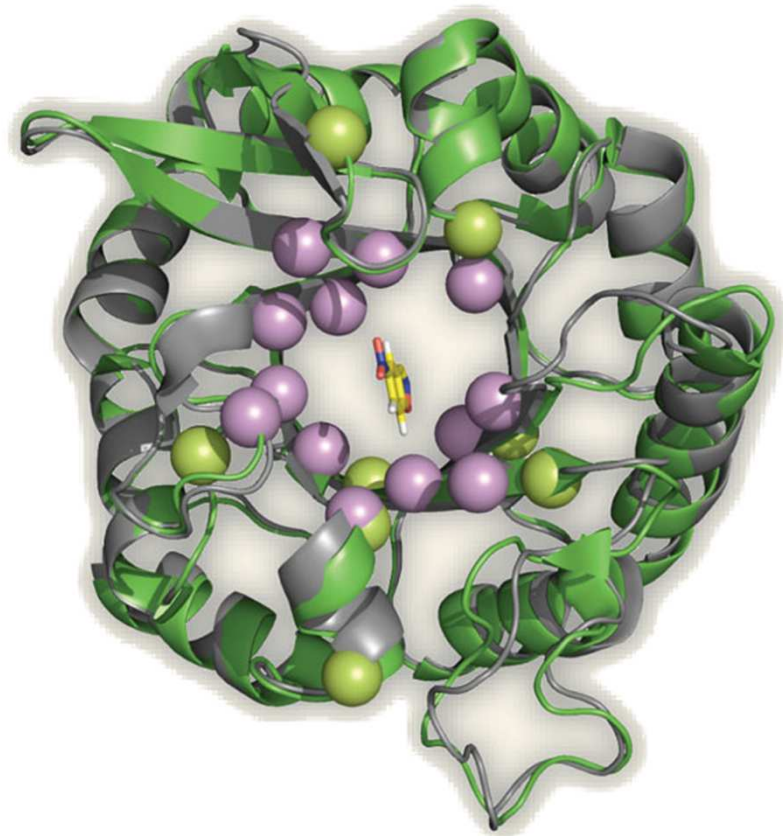


In comparison with Rh-catalyzed process,

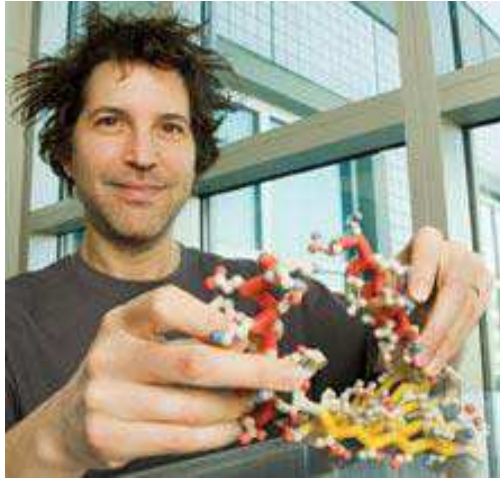
- 10-13% increase in overall yield
- 53% increase in productivity (kg/L per day)
- 19% reduction in total waste
- No need for high-pressure H₂ equipment
- Reduction in total manufacturing cost
- Elimination of all heavy metals

Contents

1. Introduction
2. Catalytic Antibodies
3. Directed Evolution
4. Computational Enzyme Design
5. Summary & Future Outlook



Computational enzyme design



David Baker

Investigator, Howard Hughes Medical Institute
Professor of Biochemistry, University of Washington
Adjunct Professor of Genome Sciences
Adjunct Professor of Physics
Adjunct Professor of Computer Science
Adjunct Professor of Chemical Engineering
Adjunct Professor of Bioengineering

1984 B.A. at Harvard University
1989 Ph.D. at University of California, Berkeley
1990-1993 Postdoctoral work at University of California, San Francisco
199X Professor of Biochemistry, University of Washington

The Baker laboratory developed the Rosetta algorithm for *de novo* protein structure prediction.

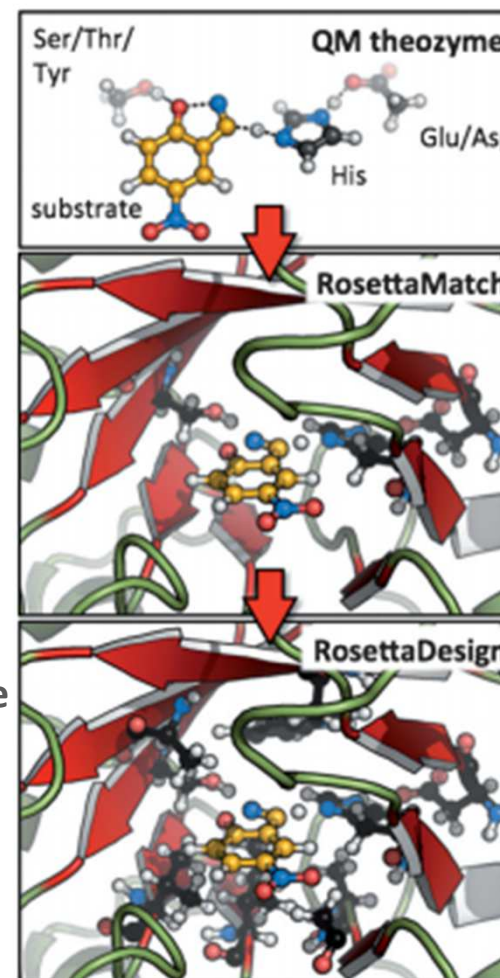
He aims to produce structural models for protein complexes as well as individual polypeptide chains.

His group is recognized as the first group to have designed a protein

Here, I'll focus on his works.

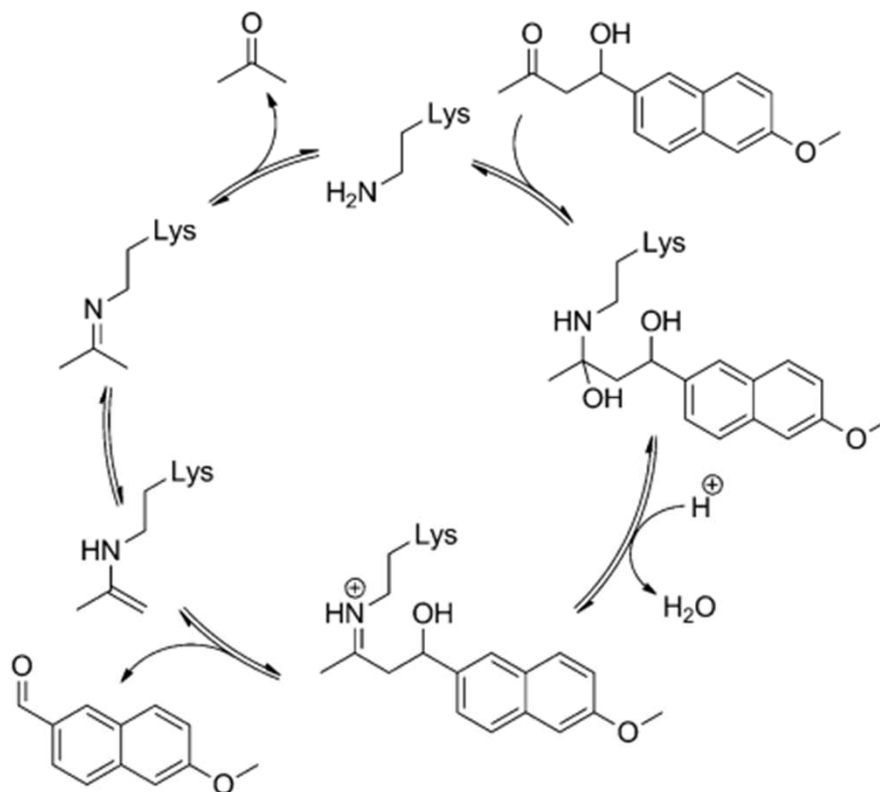
De novo computational Enzyme Design

1. QM calculation to generate “Theozyme”, functional groups stabilizing the TS.
A number of theozyme motif are usually generated.
2. Run RosettaMatch to search active sites of existing proteins for backbone position that can accommodate three-dimensional side chain rearrangement of theozyme.
3. RosettaDesign attempts to generate the best possible stabilization for a geometry .
Normally, even the highest ranked design differ quite considerably from the original theozyme geometry.
4. Final designs are assessed towards their capability to stabilize the key catalytic residues on the basis of criteria such as Rosetta energy, hydrogen bond, active site geometry etc...
5. Enzyme assay
6. Directed evolution



Achievement – Retro-Aldolases

The first example of *de novo* computational enzyme design is retro-aldolase

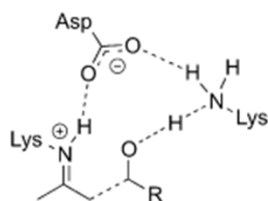


Jiang, L. *Science*, **2008**, 319, 1387

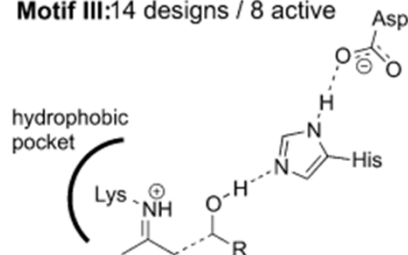
Achievement – Retro-Aldolases

Theozyme design

Motif I: 12 designs / 0 active



Motif III: 14 designs / 8 active

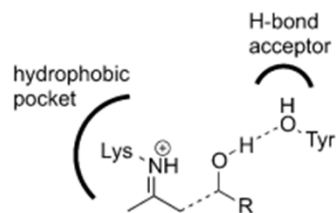


30 design showed detectable activity

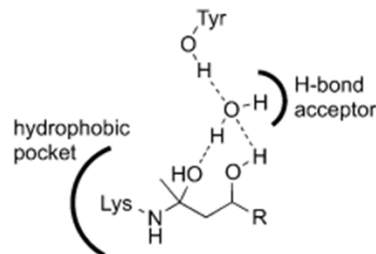
$$k_{\text{cat}}/K_M = 0.02 - 0.74\text{M}^{-1}\text{s}^{-1}$$

Enhancement of rate is inferior to that of catalytic antibodies.

Motif II: 10 designs / 0 active

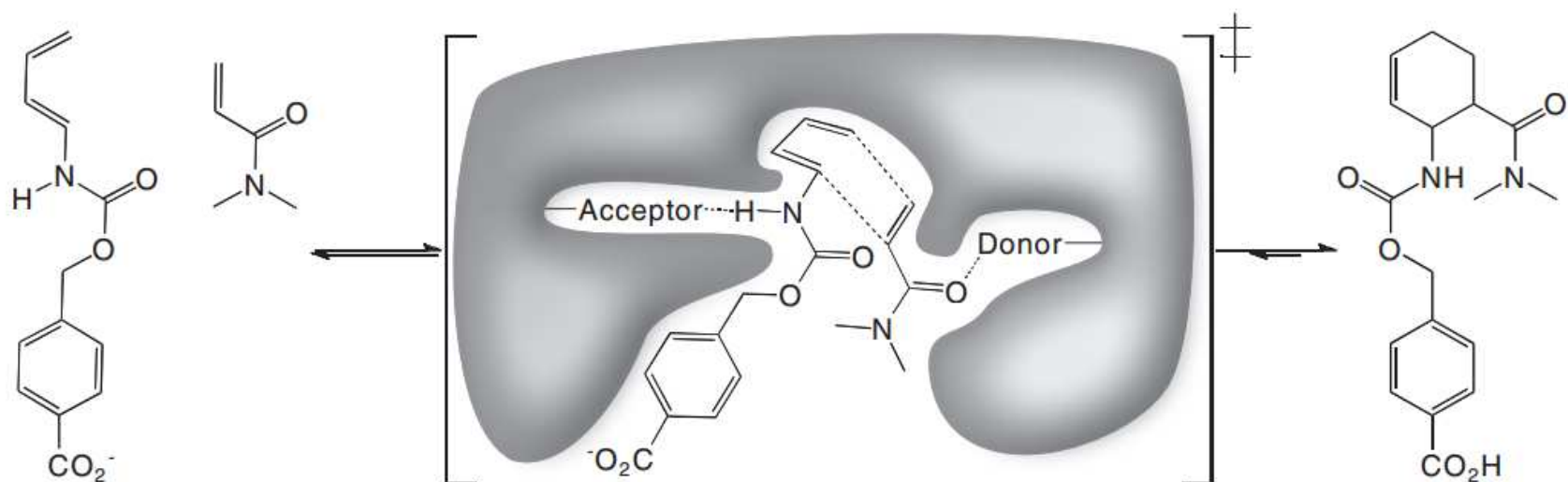


Motif IV: 38 designs / 22 active



Low activity can be attributed to dynamic distortion such as solvation, conformational flexibility.

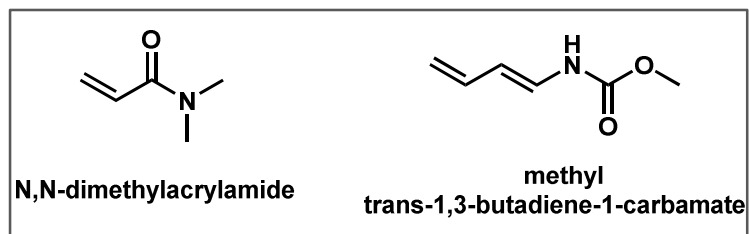
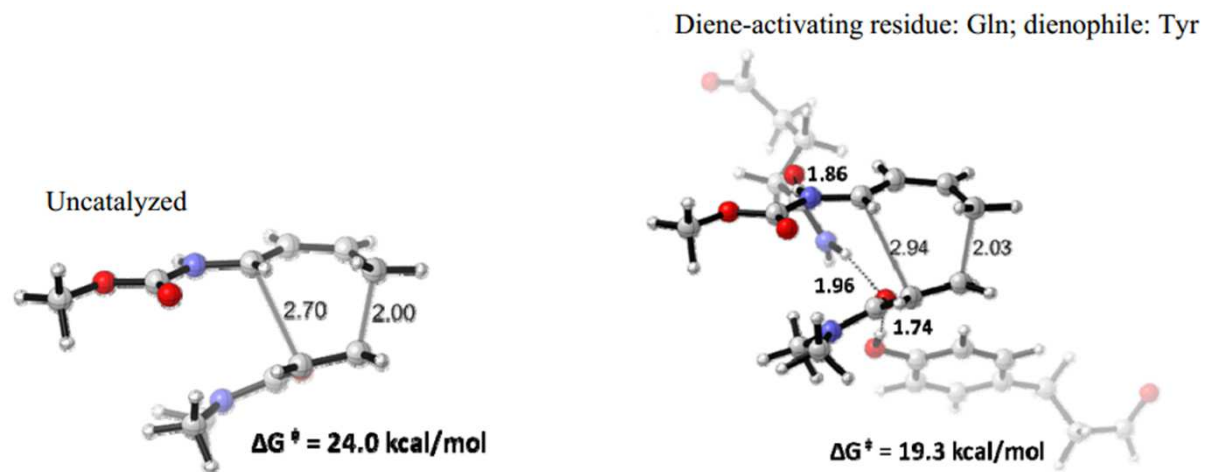
Achievement – Diels-Alderase



Designing enzyme that catalyze bimolecular bond-forming reaction is challenging.
(Both substrate must be bound in the proper relative orientation.)

In this design, hydrogen bond donor & acceptor modulate molecular orbital energies and stabilize TS to accelerate the intermolecular Diels-Alder reaction.

QM calculations



Model substrates

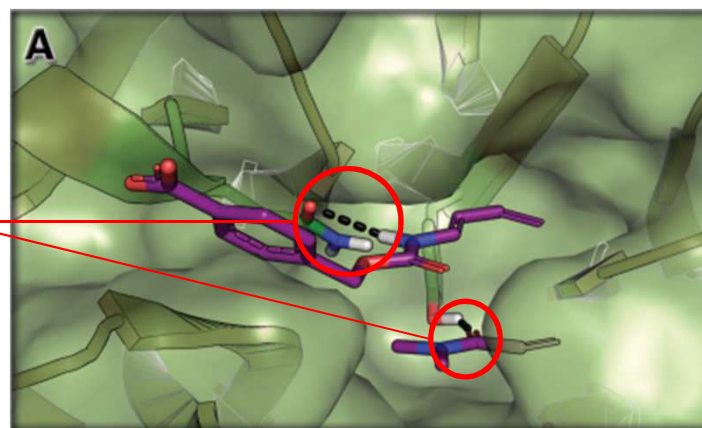
Quantum mechanical simulation predict that these hydrogen bonds can stabilize the TS by 4.7kcal/mol.

Design and screening

84 designs were synthesized through QM calculation, RosettaMatch, and RosettaDesign. Two of them were found to have Diels-Alderase activity. (DA_20_10 and DA_42_00)

Then, these Diels-Alderase were evolved to further improved the activity

Hydrogen bond



Structure of DA_20_10

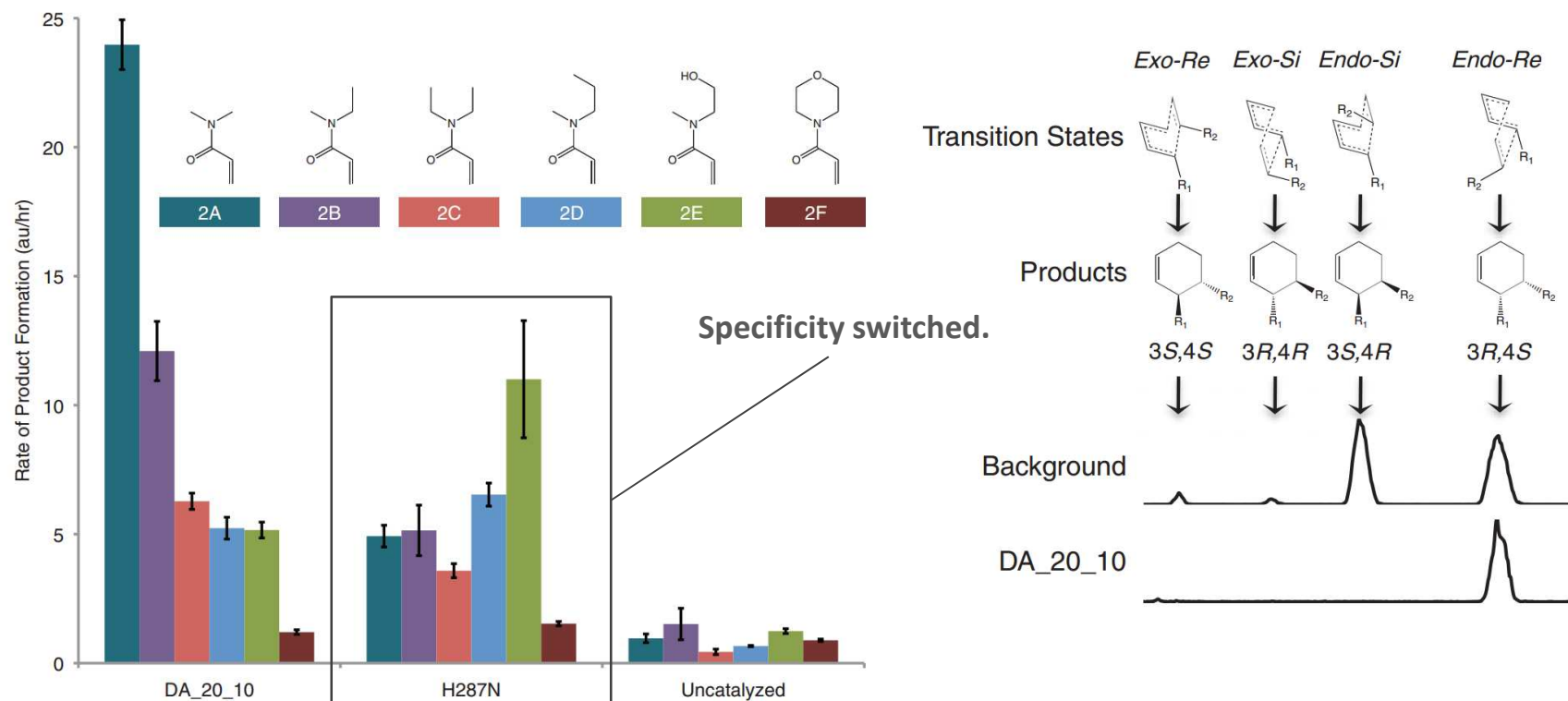
Since initial activity is promising, directed evolution can be effectively combined with computational enzyme design.

DA_20_10(100-fold active), DA_42_04(20-fold active)

Catalyst	k_{cat} (hour ⁻¹)	K_M -diene (mM)	K_M -dienophile (mM)	k_{cat}/K_M -diene (s ⁻¹ M ⁻¹)	k_{cat}/K_M -dienophile (s ⁻¹ M ⁻¹)	$k_{cat}/(K_M$ -diene × K_M -dienophile) (s ⁻¹ M ⁻¹ M ⁻¹)
DA_20_00	0.10 ± 0.02	3.5 ± 1.5	146.0 ± 2.5	0.008	0.0002	0.06
DA_20_10	2.13 ± 0.24	1.3 ± 0.1	72.8 ± 5.1	0.455	0.0081	6.23
DA_42_04	0.03 ± 0.01	0.5 ± 0.1	16.2 ± 3.2	0.017	0.0005	1.03
mAb 7D4	0.21	1.0	1.7	0.058	0.0343	20.18
mAb 4D5	0.21	1.6	5.9	0.036	0.0099	6.19

Siegel, J. B., et al. *Science*, 2010, 329, 309

Specificity and selectivity



In addition to high substrate specificity and stereoselectivity, once an initial active enzyme is engineered, it can be easily modified to catalyze similar reactions with alternate substrate.

Challenges in enzyme design

There is much room for improvement in computational enzyme design.

- Only two of 50 designs exhibited detectable activity (low success rate)
- Low activity relative to natural enzymes
- The resulting microenvironments are still far from “finely tuned”

Design through Crowd Sourcing



Structure prediction with Foldit

Pull Mode

Rank: 4 Score: 11459.528
Soloist 399: Design the Interface 6b
No bonuses or conditions

Group Competition
Soloist Competition

foldit
Solve Puzzles for Science

Cookbook

Chat - Group auto show
Chat - Puzzle auto show

CFC: aunt, can I not count, or did you have 2 additions on one and 2 on the other?
aunteen_irc: angle of shot - hold on, was 3/2
CFC: ok

Send

Chat - Global auto show
Notifications auto show

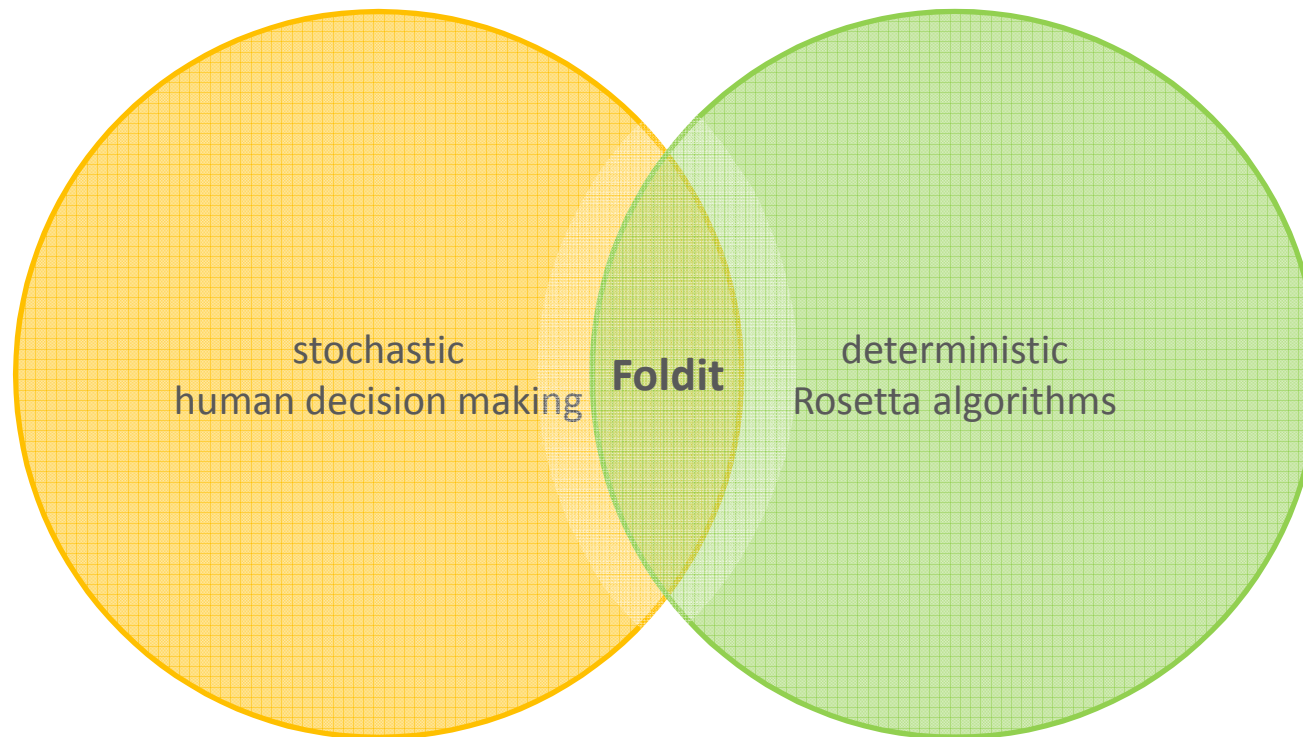
Actions Undo Social Modes Behavior View Menu

<http://fold.it/portal/>

About Foldit

Foldit is a multiplayer online game developed on the hypothesis that problem of protein structure prediction can be solved with human directed computing.

The goal of Foldit is producing accurate protein structure models through gameplay.



User interface

Tools and visualizations make the game approachable for non-scientists.

The screenshot displays the Foldit game interface. At the top, it shows 'Rank: 317' and 'Score: 2534'. Below this, a 'Group Competition' table lists group names and scores, and a 'Soloist Competition' table lists player names and scores. The main area shows a 3D protein structure with various side chains highlighted in different colors. Numbered callouts (1-12) point to specific features: 1 (interatomic repulsion), 2 (hydrogen bond), 3 (exposed hydrophobic side chain), 4 (hydrophilic side chain), 5 (high residue energy), 6 (rubber bands), 7 (freezing), 8 (score), 9 (leader board), 10 (tools and options), 11 (chat window), and 12 (cookbook). The bottom of the screen features a toolbar with icons for actions like 'Shake Sidechains', 'Wiggle', 'Freeze Protein', 'Remove Bands', 'Align Guide', 'Reset Structures', 'Reset Puzzle', 'Help', and 'Glossary'. A chat window is visible in the bottom right corner.

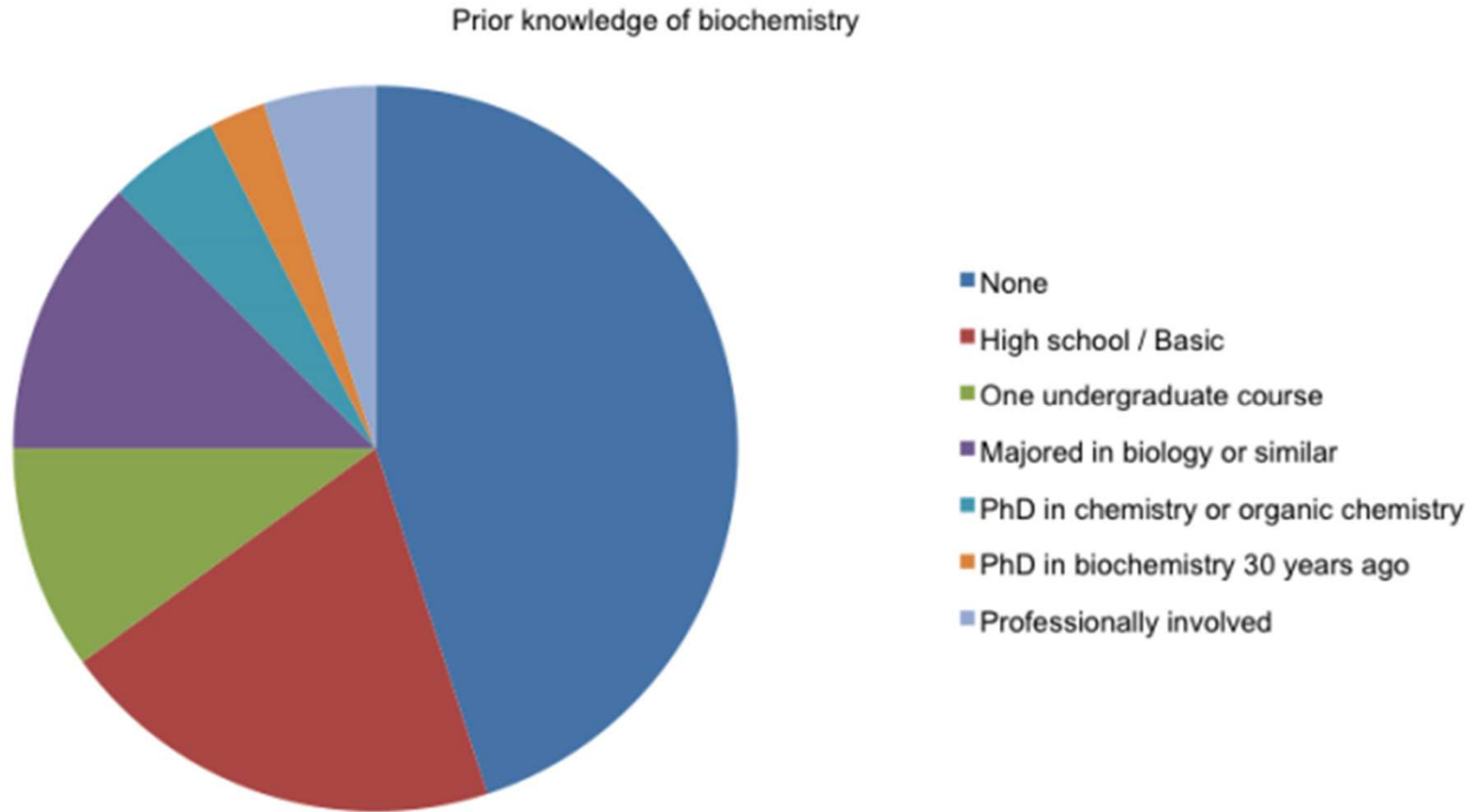
#	Group Name	Score
1	Rice Biochemistry	9174
2	Team Commonwealth	9168
3	Ukraine	9088
4	Team Canada	9085
5	Firebird BioChem	9073
6	SETI.Germany	9030
7	Bojnc.be	9001

#	Player Name	Current	Best
1	Mike Crunching For Physics	9242	9235
2	welzen	9225	9211
3	ys719	9211	9186
4	jmarkic	9211	9185
5	kevin_karplus	9186	9183
6	JINXter	9185	9183
7	eb.eric	9183	9183

1. Interatomic repulsion
2. Hydrogen bond
3. Exposed hydrophobic side chain
4. Hydrophilic side chain
5. High residue energy
6. User tool “rubber bands”
7. User tool “freezing”
8. Score
9. Leader board
10. Tools and options
11. Chat window
12. “Cookbook”

Cooper, S., et al. *Nature*, 2010, 466, 756

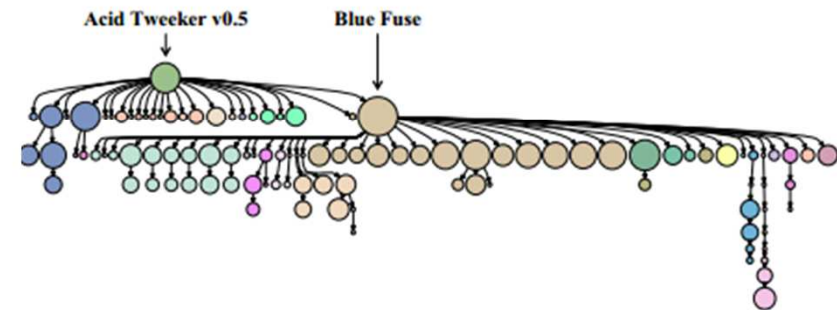
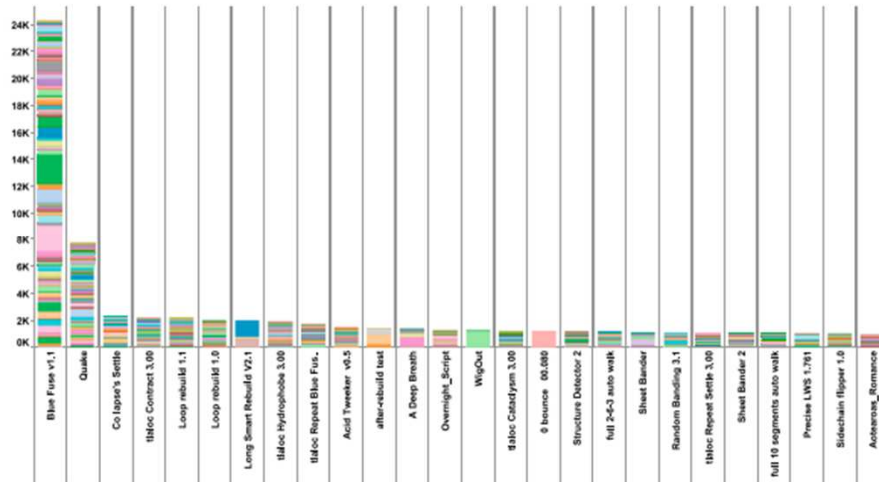
Background of players



Cooper, S., et al. *Nature*, 2010, **466**, 756

Only a minority have advanced knowledge of biochemistry

Social evolution of “recipes”



Khatib, F., *et al.* PNAS, 2011, 108, 18949

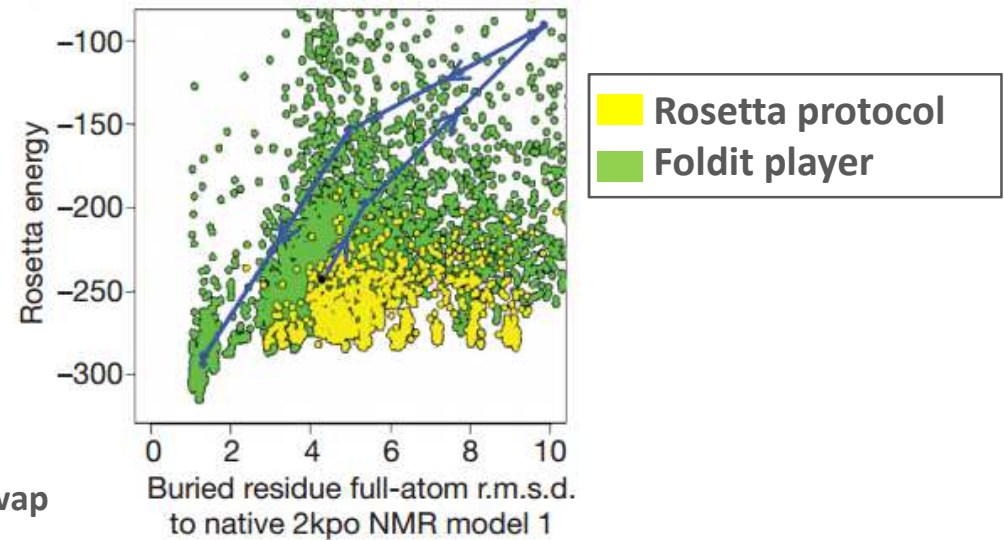
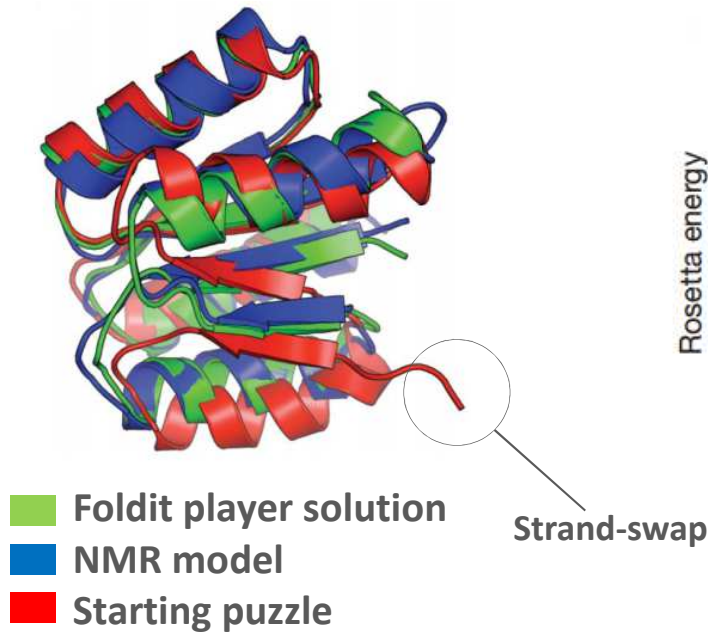
Foldit recipes are used with very different frequencies.

Popular recipes are copied and modified more often to evolve.

Thus, popular recipes spawn large numbers of descendants,

and there are multiple independent lineages each spanning many generations.

Strand-swap puzzle

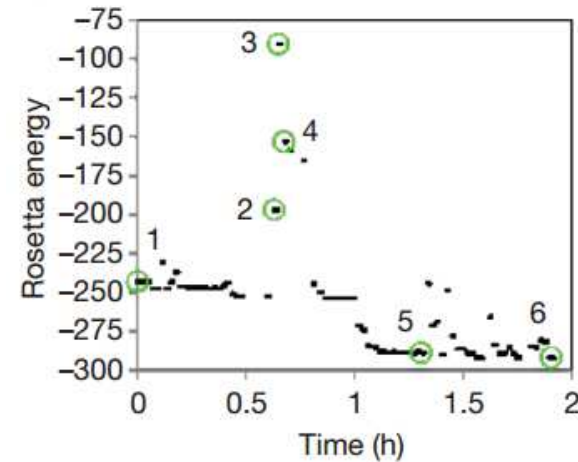
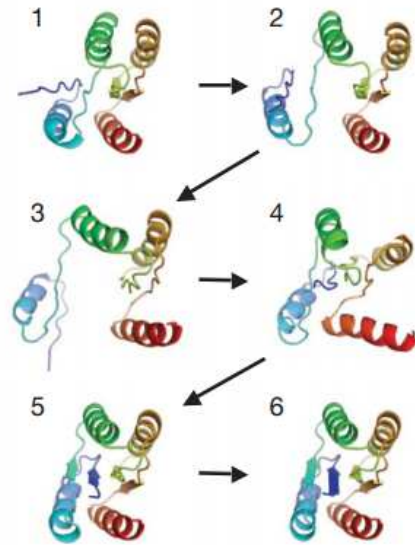


Rosetta couldn't reach swapped conformations

Humans significantly outperformed computers!

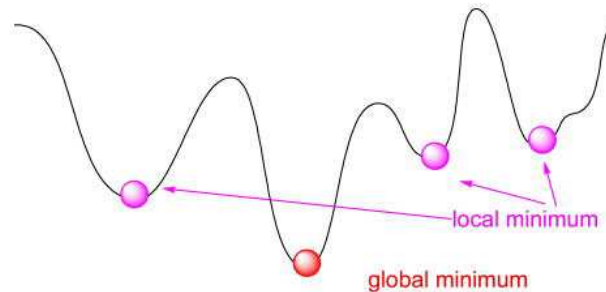
§4 Computational Enzyme Design

Key difference between humans and computers



Cooper, S., et al. *Nature*, 2010, 466, 756

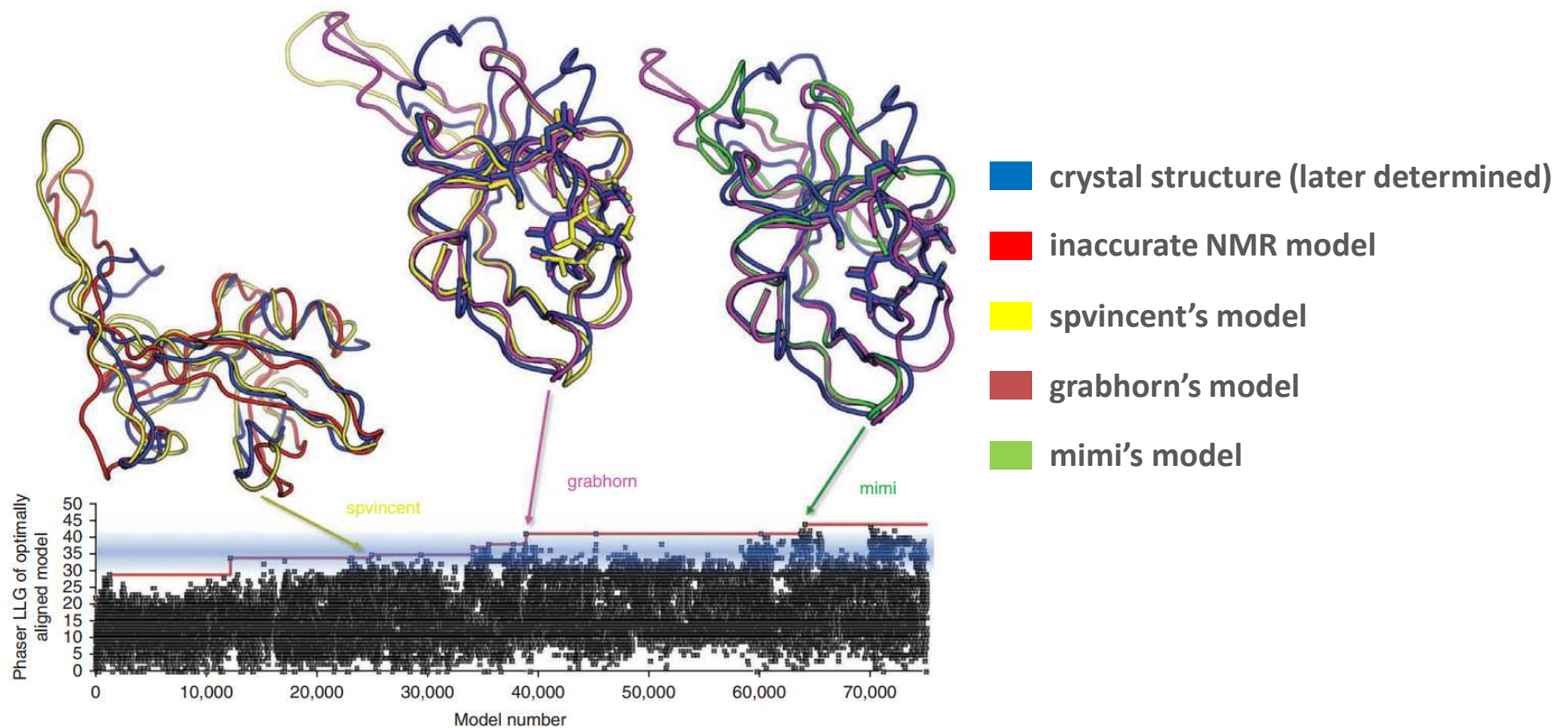
The player went through high energy conformations to reach the native state.



Deterministic minimize function has disadvantage of being trapped at local minima.

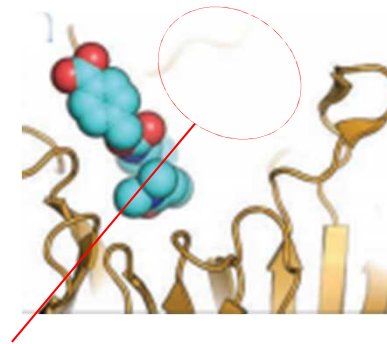
Crystal structure solved by Foldit

Crystal structure of M-PMV retroviral protease had been elusive for over a decade (Despite the availability of crystals, wide range of attempts were unsuccessful.)
Foldit players solved this long-standing protein crystal structure problem



Enzyme remodeling through Foldit

Computationally designed Diels-Alderase, DA2010 has open active site, leaving substrate exposed to solvent. To improve activity, Foldit players were enlisted to remodel loops. Players were allowed to insertions and deletions.



open active site

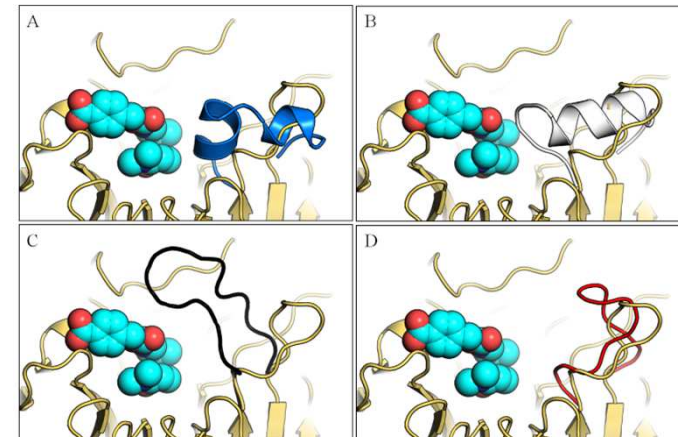
“Cover the Ligand”
Remodel any of four active site loops

After a week

4 of 69773 designs made particularly favorable interactions

Library screening

Most of variants exhibited no activity.
CE0 (based on design A) showed activity 10-fold decrease relative to DA2010



§4 Computational Enzyme Design

Enzyme remodeling through Foldit

CE0 (suboptimal interaction with the substrate or transition state)



Exploring design further

CE4 was 9-fold more active than DA2010



“Back Me Up”
Transform a loop into additional helix
to stabilize initially designed helix



After a week

Based the top design of 109,421, library was created to provide CE6
CE6 has helix-turn-helix motif, and 18-fold more active than DA2010



Helix-turn-helix motif

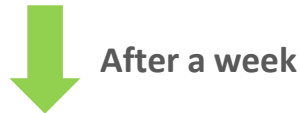
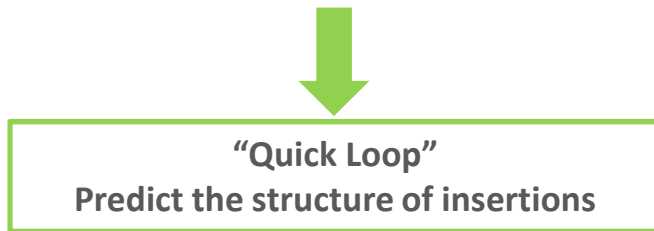
Protein	$K_{M\text{-diene}}$ (mM)	$K_{M\text{-dienophile}}$ (mM)	k_{cat} (h^{-1})	$k_{\text{cat}}/(K_{M\text{-diene}} * K_{M\text{-dienophile}})$ ($\text{s}^{-1}\text{M}^{-1}\text{M}^{-1}$)
DA2010	1.2 ± 0.2	101 ± 21	2.1 ± 0.3	4.7 ± 1.5
CE0	n.d.	n.d.	n.d.	0.5 ± 0.05
CE4	0.5 ± 0.03	31 ± 3.0	2.4 ± 0.1	42.4 ± 5.7
CE6	0.2 ± 0.03	35 ± 1.4	2.2 ± 0.1	87.3 ± 13.9

k_{uncat} under these conditions is $2.2 \times 10^{-2} \text{ M}^{-1} \text{ h}^{-1}$. n.d., not detectable.

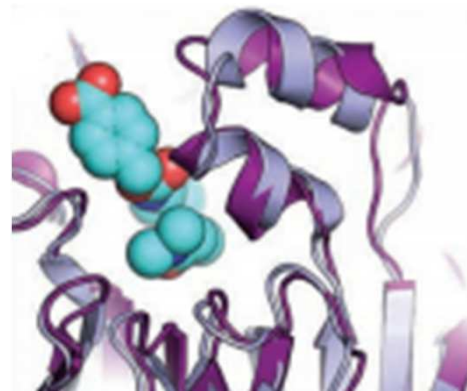
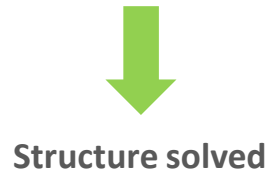
Eiben, C., et al. *Nat. Biotechnol.* **2012**, 30, 190

Enzyme remodeling through Foldit

Enhanced Diels-Alderase CE6 with community-designed helix-turn-helix motif



The lowest energy of 335,697 solutions was selected as predicted structure

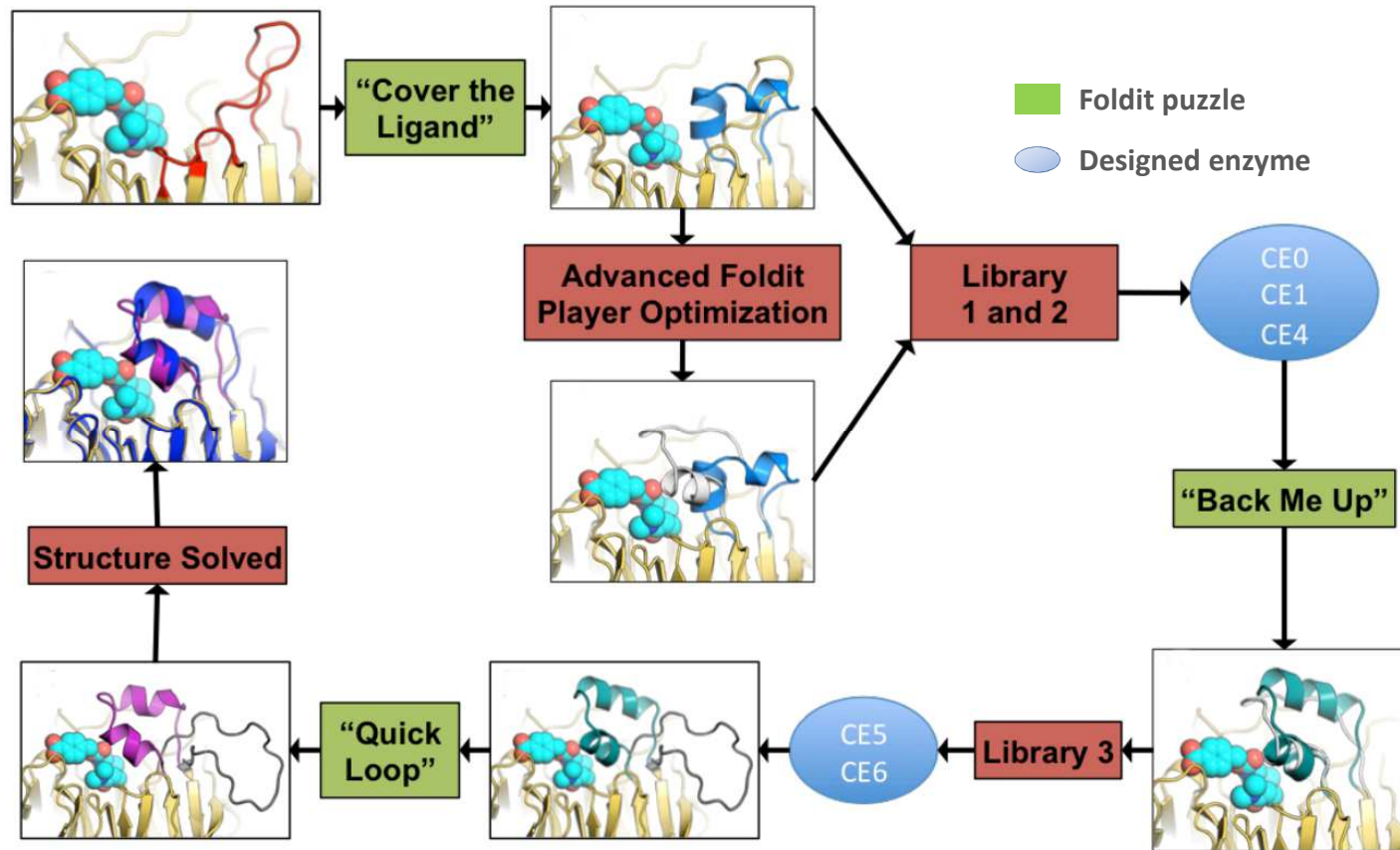


■ Predicted structure
■ Crystal structure

Eiben, C., et al. *Nat. Biotechnol.* **2012**, 30, 190

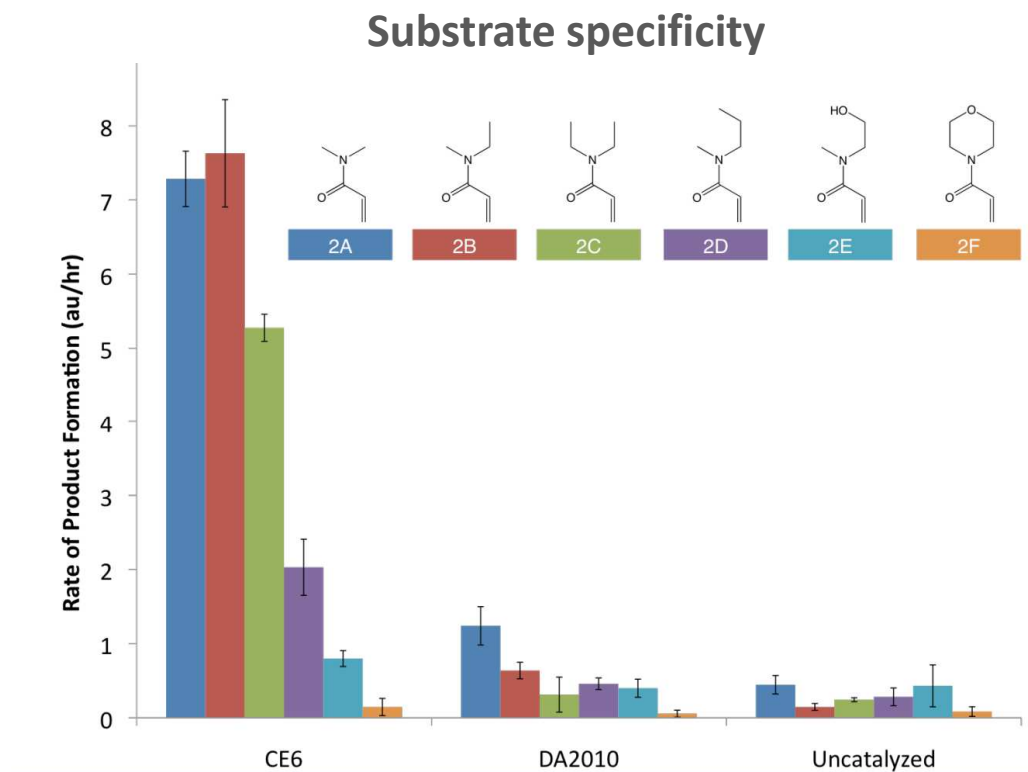
Enzyme remodeling through Foldit

Workflow of design process



Eiben, C., et al. *Nat. Biotechnol.* **2012**, 30, 190

Enzyme remodeling through Foldit



Specificity for hydrophilic dienophiles increased
However, specificity for similar-size hydrophobic dienophiles was lost
Further improvement remain possible

Predicting protein structures with a multiplayer online game

Seth Cooper¹, Firas Khatib², Adrien Treuille^{1,3}, Janos Barbero¹, Jeehyung Lee³, Michael Beenen¹, Andrew Leaver-Fay^{2,†}, David Baker^{2,4}, Zoran Popović¹ & **Foldit players**

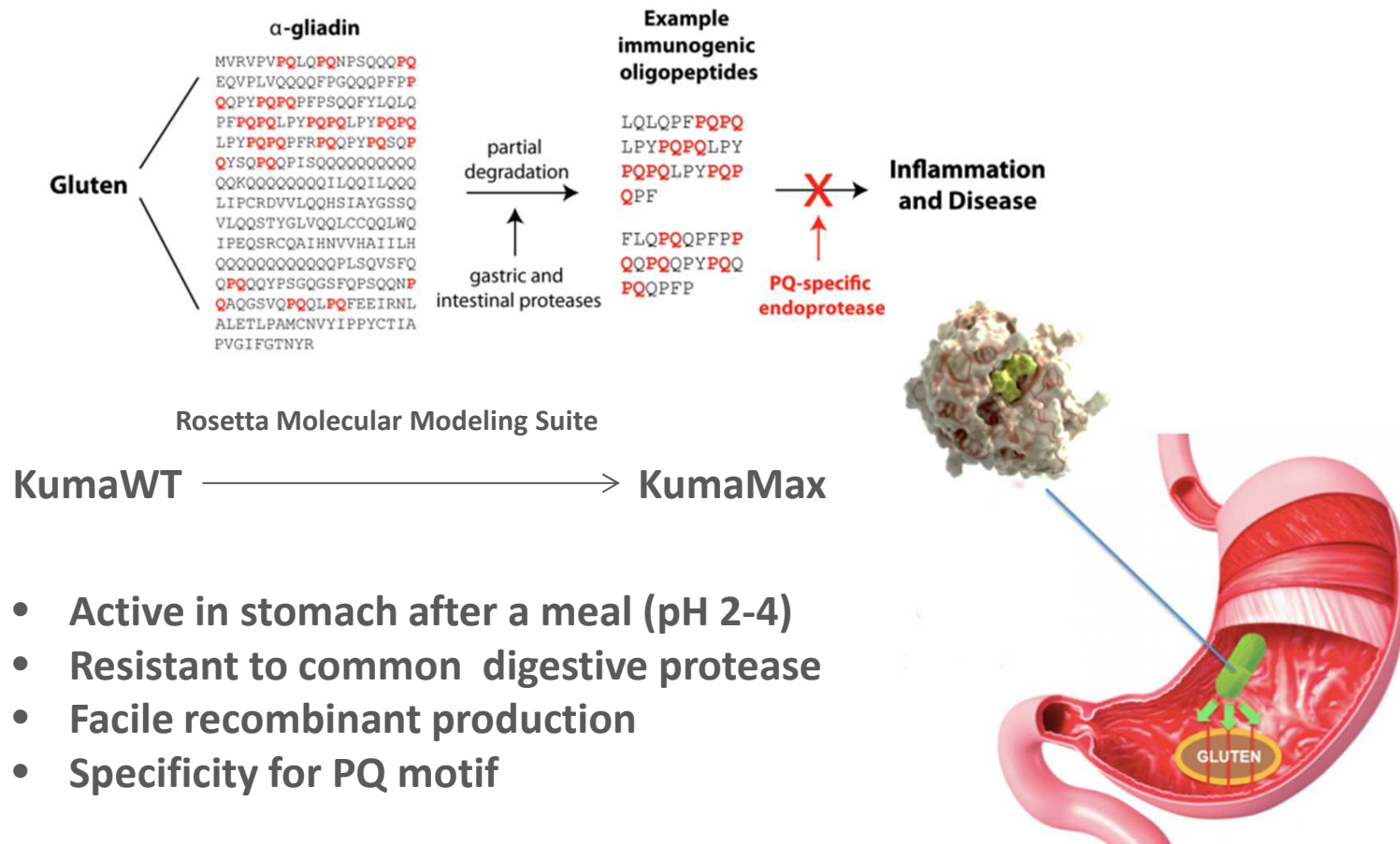
Crystal structure of a monomeric retroviral protease solved by protein folding game players

Firas Khatib¹, Frank DiMaio¹, **Foldit Contenders Group, Foldit Void Crushers Group**, Seth Cooper², Maciej Kazmierczyk³, Mirosław Gilski^{3,4}, Szymon Krzywda³, Helena Zabranska⁵, Iva Pichova⁵, James Thompson¹, Zoran Popović², Mariusz Jaskolski^{3,4} & David Baker^{1,6}

Human insight and creativity can be extended to molecular-scale design problems!

Enzyme therapy

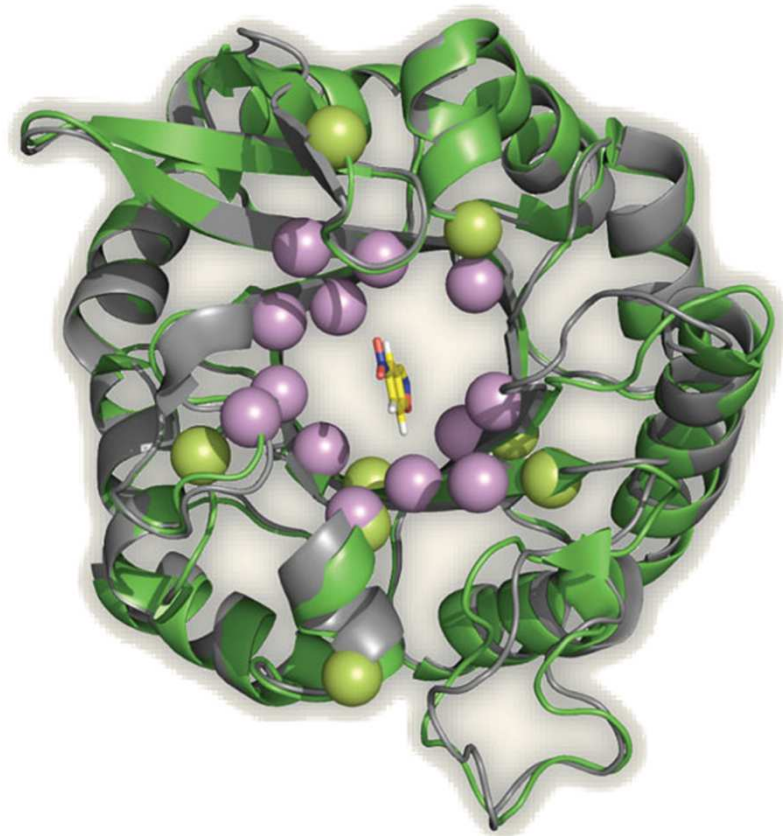
Computational design of α -gliadin peptidase as a therapeutic for celiac disease



Gordon, S. R. *J. Am. Chem. Soc.* **2012**, 134, 20513

Contents

1. Introduction
2. Catalytic Antibodies
3. Directed Evolution
4. Computational Enzyme Design
5. Summary & Future Outlook



Summary

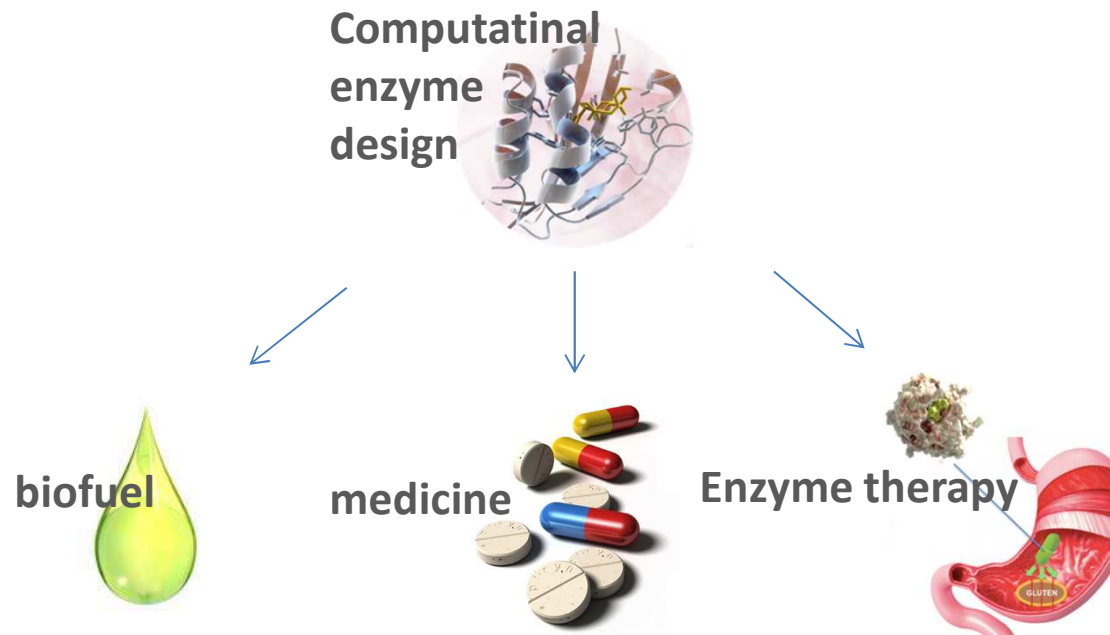
	$k_{\text{cat}}^{[a]}$	$K_M^{[b]}$	$k_{\text{cat}}/K_M^{[c]}$	$k_{\text{cat}}/k_{\text{uncat}}$	$[k_{\text{cat}}/K_M]/k_{\text{uncat}}$
nat. enzymes	av 10^5	av 10^{-4}	10^6 – 10^9	10^6 – 10^{17}	10^8 – 10^{29}
cat. antibodies	10^{-2} –1	av 10^{-4}	10^2 – 10^4	10^3 – 10^6	10^5 – 10^9
Kemp elim.					
cat. antibodies	10^{-1} –1	10^{-3} – 10^{-4}	10^2 – 10^3	10^3 – 10^6	10^7 – 10^9
comp. designs	10^{-2} –1	av 10^{-3}	10 – 10^2	10^3 – 10^6	10^7 – 10^9
evolved designs	1–20	10^{-3} – 10^{-5}	10^3 – 10^6	10^6 – 10^7	10^7 – 10^{11}
Retro-Aldol					
cat. antibodies	10^{-3} – 10^{-1}	10^{-4} – 10^{-5}	10 – 10^3	10^5 – 10^6	10^7 – 10^9
comp. designs	10^{-2} – 10^{-1}	av 10^{-4}	10^{-2} – 10^{-1}	10^3 – 10^4	10^6 – 10^7
Diels–Alder^[d]					
cat. antibodies	av 10^{-5}	av 10^{-3}	av 10	av 10^3	10^9
comp. designs	10^{-5} – 10^{-4}	10^{-1} – 10^{-4}	1– 10^2	10^3 – 10^4	10^7 – 10^{11}

[a] In units of s^{-1} . [b] In units of M. [c] In units of $M^{-1} s^{-1}$. [d] $k_{\text{cat}}/(K_{M\text{-diene}} \times K_{M\text{-dienophile}})$ instead of k_{cat}/K_M in units of $s^{-1} M^{-1} M^{-1}$ and $(k_{\text{cat}}/(K_{M\text{-diene}} \times K_{M\text{-dienophile}}))/k_{\text{uncat}}$ instead of $(k_{\text{cat}}/K_M)/k_{\text{uncat}}$ in units of $M^{-1} M^{-1}$

Computational design and directed evolution enhanced K_M value comparable to natural enzymes. However, in terms of $k_{\text{cat}}/k_{\text{uncat}}$ value, even the most active designed enzyme have rates comparable only with the least proficient of natural enzyme. Further enhancement is still required.

Future Outlook

Computational enzyme design holds promise for the production of renewable fuels, drugs, therapeutics.

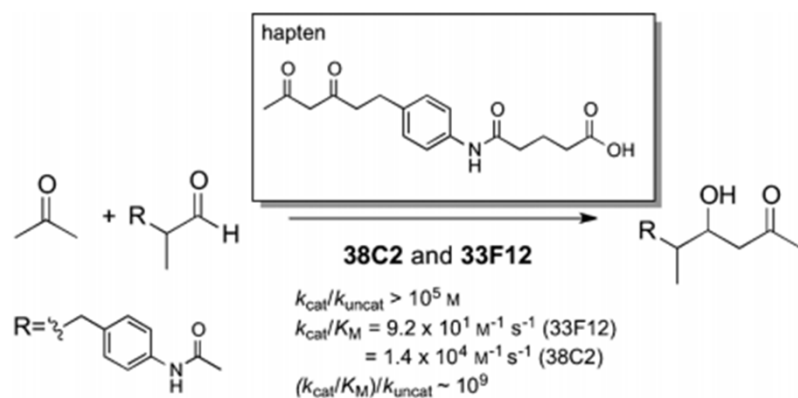


Protein engineering is one of the most dynamically developing scientific fields.

Efforts to engineer truly enzyme-like proteins are just getting started!

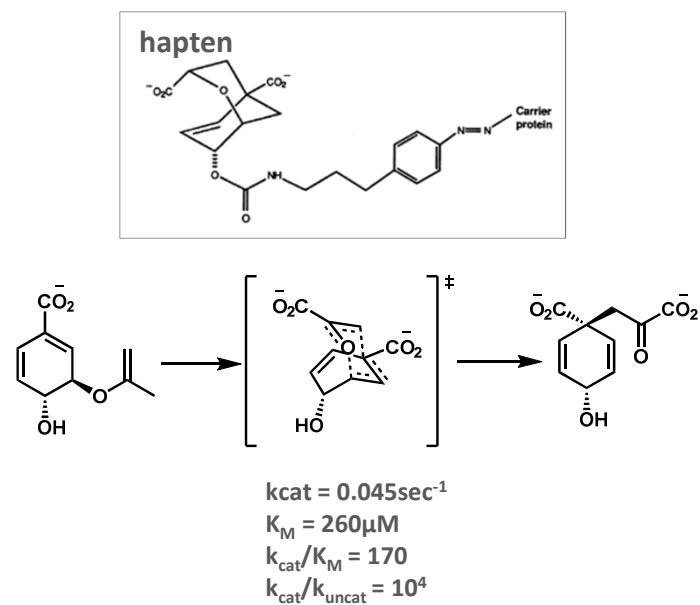
Catalytic Antibodies

Aldol reaction



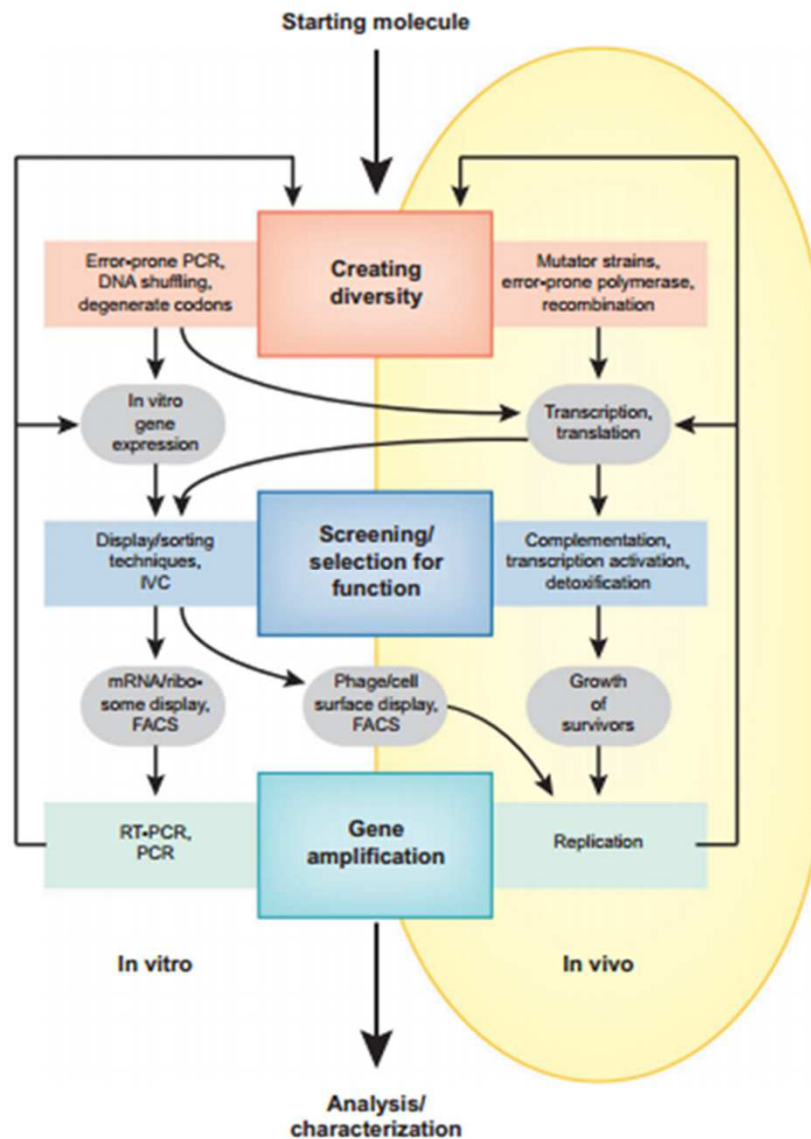
Wagner, J. *Science*, 1995, 270, 1797

Claisen rearrangement



Schultz, P. *Science*, 1988, 240, 426

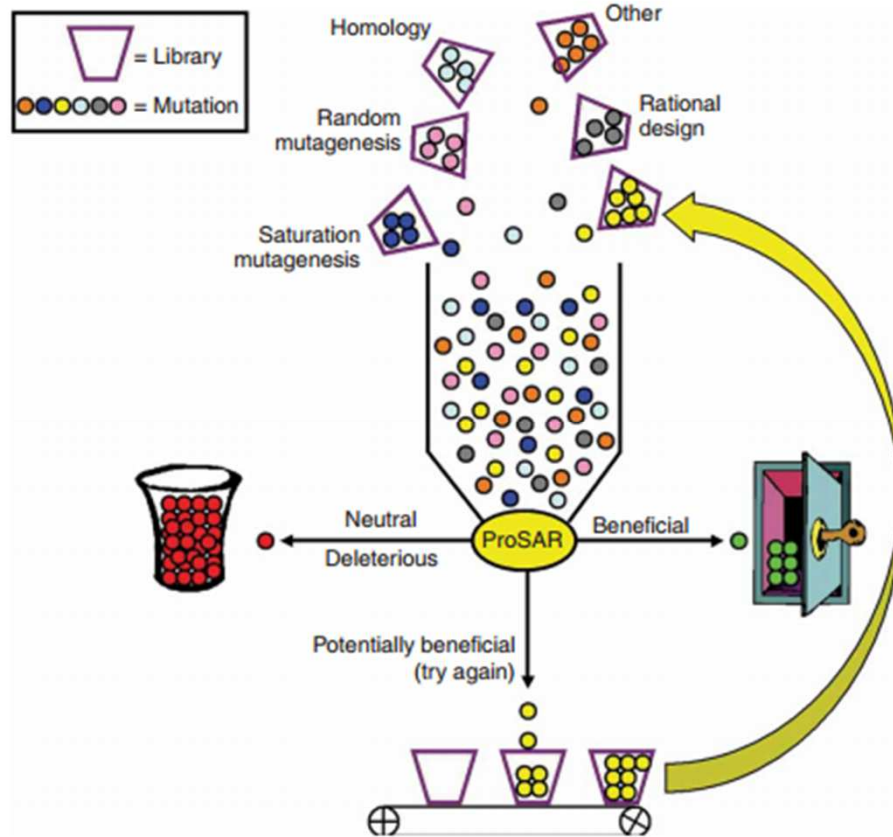
Directed evolution



Today, numerous experimental methods are available in directed evolution process.

Directed evolution

Protein sequence activity relationships (ProSAR)



Variants are categorized into four classes.

Neutral and Deleterious are discarded.

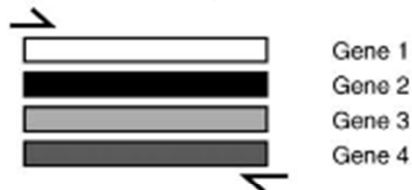
Beneficial variants will be parental enzyme in the next round.

Potentially beneficial variants are retested.

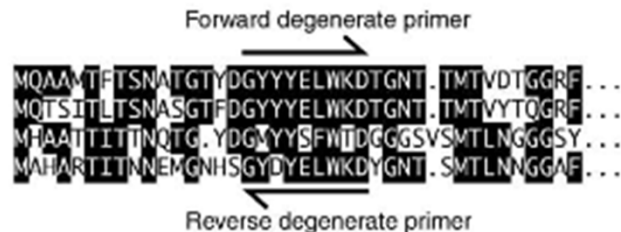
Directed evolution

Gene shuffling degenerate oligonucleotide gene shuffling(DOGS)

1. Design oligonucleotide primers with 3' ends specific for the N- or C-terminus of each candidate gene. Incorporate common nested 5' ends with suitable restriction sites for directional cloning of PCR products. PCR amplify each gene for use as PCR template.



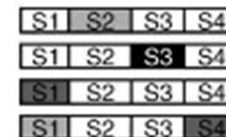
2. Design complementary degenerate primer pairs based upon one or more conserved motifs found in candidate genes.



3. Amplify each of the individual segments (S1-S4) for each gene using the degenerate primers and the common nested primers.



4. Mix segments from each gene to give desired levels of chimerisation. Regenerate full length chimeric genes by overlap extension of segments followed by PCR with primers specific for the common nested ends.



5. Digest and ligate full length fragments into an appropriate cloning vector, transform into expression host and screen individual recombinants for desired properties.